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Genetic and Genomic Testing for the Diagnosis and Management of Select Cancers and Genetic Diseases Corporate Medical Policy

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Description

Hereditary breast and ovarian cancer syndrome describe the familial cancer syndromes related to variants in the BRCA genes (BRCA1 located on chromosome 17q21, BRCA2 located on chromosome 13q12-13). The PALB2 gene is located at 16p12.2 and has 13 exons. PALB2 protein assists BRCA2 in DNA repair and tumor suppression. Families with hereditary breast and ovarian cancer syndrome have an increased susceptibility to the following types of cancer: breast cancer occurring at a young age, bilateral breast cancer, male breast cancer, ovarian cancer (at any age), cancer of the fallopian tube, primary peritoneal cancer, prostate cancer, pancreatic cancer, gastrointestinal cancers, melanoma, and laryngeal cancer.

Genetic testing is available for both those with and those at risk for various types of hereditary cancer. This review evaluates genetic testing for hereditary colorectal cancer (CRC) and polyposis syndromes, including familial adenomatous polyposis (FAP), Lynch syndrome (formerly known as hereditary nonpolyposis colorectal cancer), *MUTYH*-associated polyposis (MAP), Lynch syndrome-related endometrial cancer, juvenile polyposis syndrome (JPS), and Peutz-Jeghers syndrome (PJS).

Somatic (acquired) genetic variants in *JAK2*, *MPL*, and *CALR* genes have been implicated as the underlying molecular genetic drivers for the pathogenesis of myeloproliferative neoplasms (MPN). This evidence review addresses the use of genetic testing for *JAK2*, *MPL*, and *CALR* genes for diagnosis, prognosis, and treatment selection of patients with MPN.

In the treatment of Philadelphia chromosome-positive leukemias, various nucleic acid-based laboratory methods may be used to detect the BCR-ABL1 fusion gene for confirmation of the

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diagnosis; for quantifying mRNA *BCR-ABL1* transcripts during and after treatment to monitor disease progression or remission; and for identification of *ABL* kinase domain (KD) single nucleotide variants related to drug resistance when there is inadequate response or loss of response to tyrosine kinase inhibitors (TKIs), or disease progression.

Carrier screening is performed to identify individuals at risk of having offspring with inherited recessive single-gene disorders. Carriers are usually not at risk of developing the disease but can pass pathogenic variants to their offspring. Carrier testing may be performed in the prenatal or preconception periods.

Coding Information

Click the links below for attachments, coding tables & instructions. Attachment I - Coding Table

Policy

Germline Genetic Testing for Hereditary Breast/Ovarian Cancer Syndrome and Other High-Risk Cancers (BRCA1, BRCA2, PALB2)

Individuals With Cancer or With a Personal History of Cancer

Genetic testing for *BRCA1*, *BRCA2*, and *PALB2* variants in cancer-affected individuals may be considered **medically necessary** under any of the following circumstances:

- Individuals with any close blood relative with a known *BRCA1*, *BRCA2*, or *PALB2* pathogenic/likely pathogenic variant (see Policy Guidelines for definitions and for testing strategy).
- Individuals meeting the criteria below but with previous limited testing (eg, single gene and/or absent deletion duplication analysis)
- Personal history of breast cancer and 1 or more of the following:
 - Diagnosed at age ≤45 years; or
 - Diagnosed at age 46 to 50 years with:
 - An additional breast cancer primary at any age; or
 - ≥1 close relative (see Policy Guidelines) with breast, ovarian, pancreatic, or prostate cancer at any age; or
 - An unknown or limited family history
 - o Diagnosed at age ≤60 years with:
 - Triple-negative breast cancer; or
 - Diagnosed at any age with:
 - ≥1 close blood relative with:
 - Breast cancer diagnosed at age ≤50 years; or
 - Ovarian carcinoma; or
 - Metastatic or intraductal/cribriform prostate cancer, or high-risk group or very-high-risk group (see Policy Guidelines) prostate cancer; or

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- Pancreatic cancer: or
- ≥3 total diagnoses of breast cancer in individual and/or close blood relatives; or
- Ashkenazi Jewish ancestry
- Diagnosed at any age with male breast cancer
- Personal history of epithelial ovarian carcinoma (including fallopian tube cancer or peritoneal cancer) at any age
- Personal history of exocrine pancreatic cancer at any age
- Personal history of metastatic or intraductal/cribriform histology prostate cancer at any age; or high-risk group or very-high-risk group prostate cancer at any age
- Personal history of prostate cancer at any age with:
 - ≥1 close blood relative with ovarian carcinoma, pancreatic cancer, or metastatic or intraductal/cribriform prostate cancer at any age, or breast cancer at age ≤50 years; or
 - ≥2 close blood relatives with breast or prostate cancer (any grade) at any age;
 or
 - Ashkenazi Jewish ancestry
- Personal history of a *BRCA1*, *BRCA2*, or *PALB2* pathogenic/likely pathogenic variant identified on tumor genomic testing that has clinical implications if also identified in the germline.

Individuals Without Cancer or With Other Personal History of Cancer

Genetic testing for *BRCA1*, *BRCA2*, and *PALB2* variants of cancer-unaffected individuals and individuals with cancer but not meeting the above criteria (including individuals with cancers unrelated to hereditary breast and ovarian cancer syndrome) may be considered **medically necessary** under any of the following circumstances:

- An individual with or without cancer and not meeting the above criteria but who has a
 1st- or 2nd-degree blood relative meeting any criterion listed above for Patients With
 Cancer (except individuals who meet criteria only for systemic therapy decisionmaking). If the individual with cancer has pancreatic cancer or prostate cancer
 (metastatic or intraductal/cribriform or high-risk group or very-high-risk group) then
 only first-degree relatives should be offered testing unless there are other family
 history indications for testing.
- An individual with any type of cancer (cancer related to hereditary breast and ovarian cancer syndrome but not meeting above criteria, or cancer unrelated to hereditary breast and ovarian cancer syndrome) or unaffected individual who otherwise does not meet the criteria above but has a probability >5% of a *BRCA1/2* or *PALB2* pathogenic variant based on prior probability models (eg, Tyrer-Cuzick, BRCAPro, Pennll).

See Policy Guidelines: Testing Unaffected Individuals.

Genetic testing for *BRCA*1 and *BRCA*2 variants of cancer-affected individuals or cancer-unaffected individuals with a family history of cancer when criteria above are not met is considered **investigational**.

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Testing for *PALB2* variants in individuals who do not meet the criteria outlined above is considered **investigational**.

Genetic testing in minors for *BRCA1*, *BRCA2*, and *PALB2* variants for hereditary breast and ovarian cancer syndrome is considered **investigational** (see Policy Guidelines).

Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes

APC Testing

Genetic testing of the APC gene may be considered **medically necessary** in the following individuals:

- At-risk relatives (see Policy Guidelines section) of individuals with familial adenomatous polyposis (FAP) and/or a known APC variant.
- Individuals with a differential diagnosis of attenuated FAP versus MUTYH-associated polyposis (MAP) versus Lynch syndrome. Whether testing begins with APC variants or screening for mismatch repair (MMR) variants depends on clinical presentation.

Genetic testing for APC gene variants is considered **investigational** for colorectal cancer (CRC) individuals with classical FAP for confirmation of the FAP diagnosis.

Testing for germline *APC* gene variants for inherited CRC syndromes is considered **investigational** in all other situations.

MUTYH Testing

Genetic testing of the MUTYH gene may be considered **medically necessary** in the following individuals:

• Individuals with a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome and a negative result for APC gene variants. A family history of no parents or children with FAP is consistent with MAP (autosomal recessive).

Testing for germline *MUTYH* gene variants for inherited CRC syndromes is considered **investigational** in all other situations.

MMR Gene Testing

Genetic testing of MMR genes (MLH1, MSH2, MSH6, PMS2) may be considered **medically necessary** in the following individuals:

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- Individuals with CRC with tumor testing suggesting germline MMR deficiency or meeting clinical criteria for Lynch syndrome (see Policy Guidelines section).
- Individuals with endometrial cancer with tumor testing suggesting germline MMR deficiency or meeting clinical criteria for Lynch syndrome (see Policy Guidelines section).
- At-risk relatives (see Policy Guidelines section) of individuals with Lynch syndrome with a known pathogenic/likely pathogenic MMR gene variant.
- Individuals with a differential diagnosis of attenuated FAP versus MAP versus Lynch syndrome. Whether testing begins with APC variants or screening for MMR genes depends on clinical presentation.
- Individuals without CRC but with a family history meeting the Amsterdam or Revised Bethesda criteria, or documentation of 5% or higher predicted risk of the syndrome on a validated risk prediction model (e.g. MMRpro, PREMM5 or MMRpredict), when no affected family members have been tested for MMR variants.

Testing for germline MMR gene variants for inherited CRC syndromes is considered **investigational** in all other situations.

EPCAM Testing

Genetic testing of the *EPCAM* gene may be considered **medically necessary** when any 1 of the following 3 major criteria (solid bullets) is met:

- Individuals with CRC, for the diagnosis of Lynch syndrome (see Policy Guidelines section) when:
 - Tumor tissue shows lack of MSH2 protein expression by immunohistochemistry and individual is negative for an MSH2 germline variant; OR
 - Tumor tissue shows a high level of microsatellite instability and individual is negative for a germline variant in MLH1, MSH2, MSH6, and PMS2; OR
- At-risk relatives (see Policy Guidelines section) of individuals with Lynch syndrome with a known pathogenic/likely pathogenic *EPCAM* variant; OR
- Individuals without CRC but with a family history meeting the Amsterdam or Revised Bethesda criteria, or documentation of 5% or higher predicted risk of the syndrome on a validated risk prediction model (e.g. MMRpro, PREMM5 or MMRpredict), when no affected family members have been tested for MMR variants, and when sequencing for MMR variants is negative.

Testing for germline *EPCAM* gene variants for inherited CRC syndromes is considered **investigational** in all other situations.

BRAF V600E or MLH1 promoter methylation

Somatic genetic testing for *BRAF* V600E or *MLH1* promoter methylation may be considered **medically necessary** to exclude a diagnosis of Lynch syndrome when the MLH1 protein is not expressed in a CRC tumor on immunohistochemical analysis.

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Testing for somatic *BRAF V600E* or *MLH1* promoter methylation to exclude a diagnosis of Lynch syndrome is considered **investigational** in all other situations.

SMAD4 and BMPR1A Testing

Genetic testing of SMAD4 and BMPR1A genes may be considered **medically necessary** when any 1 of the following major criteria (solid bullets) is met:

- Individuals with a clinical diagnosis of juvenile polyposis syndrome based on the presence of any 1 of the following:
 - o at least 5 juvenile polyps in the colon
 - o multiple juvenile polyps found throughout the gastrointestinal tract
 - any number of juvenile polyps in a person with a known family history of juvenile polyps.
- At-risk relative of an individual suspected of or diagnosed with juvenile polyposis syndrome.

Testing for germline SMAD4 and BMPR1A gene variants for inherited CRC syndromes is considered **investigational** in all other situations.

STK11 Testing

Genetic testing for *STK11* gene variants may be considered **medically necessary** when any 1 of the following major criteria (solid bullets) is met:

- Individuals with a clinical diagnosis of Peutz-Jeghers syndrome based on the presence of any 2 of the following:
 - presence of 2 or more histologically confirmed Peutz-Jeghers polyps of the gastrointestinal tract.
 - characteristic mucocutaneous pigmentation of the mouth, lips, nose, eyes, genitalia, or fingers
 - o family history of Peutz-Jeghers syndrome.
- At-risk relative of an individual suspected of or diagnosed with Peutz-Jeghers syndrome.

Testing for germline *STK11* gene variants for inherited CRC syndromes is considered **investigational** in all other situations.

Other Variants

Genetic testing of all other genes for an inherited CRC syndrome is considered **investigational**.

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Genetic Counseling

Pre- and post-test genetic counseling may be considered **medically necessary** as an adjunct to the genetic testing itself.

JAK2, MPL, and CALR Testing for Myeloproliferative Neoplasms

JAK2 testing may be considered **medically necessary** in the diagnosis of individuals presenting with clinical, laboratory, or pathologic findings suggesting polycythemia vera, essential thrombocythemia (ET), or primary myelofibrosis (PMF). Based on criteria from the World Health Organization and the International Consensus Classification for diagnosis of PV, documentation of a serum erythropoietin level below the reference range for normal is recommended before JAK2 testing (See Policy Guidelines).

MPL and CALR testing may be considered **medically necessary** in the diagnosis of individuals presenting with clinical, laboratory, or pathologic findings suggesting ET or PMF.

JAK2, MPL, and CALR testing is considered **investigational** in all other circumstances including, but not limited to, the following situations:

- Diagnosis of nonclassic forms of myeloproliferative neoplasms (MPNs)
- Molecular phenotyping of individuals with MPNs
- Monitoring, management, or selecting treatment in individuals with MPNs.

BCR-ABL1 Testing in Chronic Myelogenous Leukemia and Acute Lymphoblastic Leukemia

Chronic Myelogenous Leukemia

BCR-ABL1 qualitative testing for the presence of the fusion gene may be considered **medically necessary** for the diagnosis of chronic myeloid leukemia (see Policy Guidelines section).

BCR-ABL1 testing for messenger RNA transcript levels by quantitative real-time reverse transcription-polymerase chain reaction at baseline before initiation of treatment and at appropriate intervals (see Policy Guidelines section) may be considered **medically necessary** for monitoring of chronic myeloid leukemia treatment response and remission.

Evaluation of *ABL* kinase domain (KD) single nucleotide variants to assess individuals for tyrosine kinase inhibitor resistance may be considered **medically necessary** when there is an inadequate initial response to treatment or any sign of loss of response (see Policy Guidelines section); and/or when there is a progression of the disease to the accelerated or blast phase.

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Evaluation of *ABL* KD single nucleotide variants is considered **investigational** for monitoring in advance of signs of treatment failure or disease progression.

Acute Lymphoblastic Leukemia

BCR-ABL1 testing for messenger RNA transcript levels by quantitative real-time reverse transcription-polymerase chain reaction at baseline before initiation of treatment and at appropriate intervals during therapy (see Policy Guidelines section) may be considered **medically necessary** for monitoring of Philadelphia chromosome-positive acute lymphoblastic leukemia treatment response and remission.

Evaluation of *ABL* KD single nucleotide variants to assess individuals for tyrosine kinase inhibitor resistance may be considered **medically necessary** when there is an inadequate initial response to treatment or any sign of loss of response.

Evaluation of *ABL* KD single nucleotide variants is considered **investigational** for monitoring in advance of signs of treatment failure or disease progression.

Carrier Screening for Genetic Diseases

Targeted Risk-Based Carrier Screening

Targeted carrier screening for X-linked and autosomal recessive genetic diseases is considered **medically necessary** for individuals who are pregnant or are considering pregnancy and are at increased risk of having offspring with an X-linked or autosomal recessive disease when one of the following criteria is met:

- One or both individuals have a first- or second-degree relative who is affected; OR
- One individual is known to be a carrier; OR
- One or both individuals are members of a population known to have a carrier rate that exceeds a threshold considered appropriate for testing for a particular condition.

AND all of the following criteria are met:

- The natural history of the disease is well understood and there is a reasonable likelihood that the disease is one with high morbidity or early mortality in the homozygous or compound heterozygous state (see Policy Guidelines);
- Alternative biochemical or other clinical tests to definitively diagnose carrier status
 are not available, or, if available, provide an indeterminate result or are individually
 less efficacious than genetic testing;
- The genetic test has adequate clinical validity to guide clinical decision-making and residual risk is understood;
- An association of the marker with the disorder has been established;

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- If targeted testing is performed by a panel, the panel meets the minimum number of recommended gene variants but does not exceed the maximum, as determined by professional clinical guidelines (see Policy Guidelines). Non-targeted panels can be used instead of targeted testing when the criteria for non-targeted carrier screening are met (see below);
- Previous carrier screening or individual targeted gene testing for the gene variant(s) of interest has not been performed (see Policy Guidelines).

All targeted carrier screening not meeting any of the above criteria is considered **investigational**.

First-degree relatives include a biological parent, brother, sister, or child; second-degree relatives include a biologic grandparent, aunt, uncle, niece, nephew, grandchildren, and half-sibling.

Non-Targeted Carrier Screening

Non-targeted carrier screening panels for autosomal recessive and X-linked genetic disorders may be considered **medically necessary** as an alternative to testing of individual genes (eg, *SMN1* gene and *CFTR* gene) for individuals who are pregnant or are considering pregnancy at any risk level including high risk and average risk when all of the following criteria are met:

- The natural history of each disease is well understood and there is reasonable likelihood that the disease is one with high morbidity or early mortality in the homozygous or compound homozygous state (see Policy Guidelines);
- Alternative biochemical or other clinical tests to definitively diagnose carrier status
 are not available, or, if available, provide an indeterminate result or are individually
 less efficacious than genetic testing;
- The genetic test has adequate clinical validity to guide clinical decision-making and residual risk is understood;
- An association of the markers with the disorders has been established:
- If testing is performed by a panel, the panel meets the minimum number of recommended gene variants but does not exceed the maximum, as determined by professional clinical guidelines (see Policy Guidelines);
- Previous carrier screening has not been performed (see Policy Guidelines).

Non-targeted carrier screening panels are considered **investigational** in all other situations when above criteria are not met (see Policy Guidelines).

Summary of Evidence

Germline Genetic Testing for Hereditary Breast/Ovarian Cancer Syndrome and Other High-Risk Cancers (BRCA1, BRCA2, PALB2)

For individuals who have cancer or a personal or family cancer history and meet criteria suggesting a risk of hereditary breast and ovarian cancer (HBOC) syndrome who receive

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genetic testing for a *BRCA1* or *BRCA2* variant, the evidence includes a TEC Assessment and studies of variant prevalence and cancer risk. Relevant outcomes are overall survival (OS), disease-specific survival, test validity, and quality of life (QOL). The accuracy of variant testing has been shown to be high. Studies of lifetime risk of cancer for carriers of a *BRCA* variant have shown a risk as high as 85%. Knowledge of *BRCA* variant status in individuals at risk of a *BRCA* variant may impact health care decisions to reduce risk, including intensive surveillance, chemoprevention, and/or prophylactic intervention. In individuals with *BRCA1* or *BRCA2* variants, prophylactic mastectomy and oophorectomy have been found to significantly increase disease-specific survival and OS. Knowledge of *BRCA* variant status in individuals diagnosed with breast cancer may impact treatment decisions. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have other high-risk cancers (eg, cancers of the fallopian tube, pancreas, prostate) who receive genetic testing for a *BRCA1* or *BRCA2* variant, the evidence includes studies of variant prevalence and cancer risk. Relevant outcomes are OS, disease-specific survival, test validity, and QOL. The accuracy of variant testing has been shown to be high. Knowledge of *BRCA* variant status in individuals with other high-risk cancers can inform decisions regarding genetic counseling, chemotherapy, and enrollment in clinical trials. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals with a risk of HBOC syndrome who receive genetic testing for a PALB2 variant, the evidence includes studies of clinical validity and studies of breast cancer risk, including a meta-analysis. Relevant outcomes are OS, disease-specific survival, and test validity. Evidence supporting clinical validity was obtained from numerous studies reporting relative risks (RRs) or odds ratios (ORs). Study designs included family segregation, kin-cohort, familybased case-control, and population-based case-control. The number of pathogenic variants identified in studies varied from 1 (founder mutations) to 48. The RR for breast cancer associated with a PALB2 variant ranged from 2.3 to 13.4, with the 2 family-based studies reporting the lowest values. Evidence of preventive interventions in women with PALB2 variants is indirect, relying on studies of high-risk women and BRCA carriers. These interventions include screening with magnetic resonance imaging, chemoprevention, and risk-reducing mastectomy. Given the penetrance of PALB2 variants, the outcomes following bilateral and contralateral risk-reducing mastectomy examined in women with a family history consistent with hereditary breast cancer (including BRCA1 and BRCA2 carriers) can be applied to women with PALB2 variants, with the benefit-to-risk balance affected by penetrance. In women at high-risk of hereditary breast cancer who would consider riskreducing interventions, identifying a PALB2 variant provides a more precise estimated risk of developing breast cancer compared with family history alone and can offer women a more accurate understanding of benefits and potential harms of any intervention. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes
For individuals who are suspected of attenuated familial adenomatous polyposis
(FAP), MUTYH-associated polyposis (MAP), and Lynch syndrome who receive genetic testing

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for adenomatous polyposis coli (*APC*), or are at-risk relatives of patients with FAP who receive genetic testing for *MUTYH* after a negative *APC* test result, the evidence includes a TEC Assessment. Relevant outcomes are overall survival (OS), disease-specific survival, and test accuracy and validity. For patients with an *APC* variant, enhanced surveillance and/or prophylactic treatment will reduce the future incidence of colon cancer and improve health outcomes. A related familial polyposis syndrome, MAP syndrome, is associated with variants in the *MUTYH* gene. Testing for this genetic variant is necessary when the differential diagnosis includes both FAP and MAP because distinguishing between the 2 leads to different management strategies. Depending on the presentation, Lynch syndrome may be part of the same differential diagnosis. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who (1) are suspected of attenuated FAP, MAP, and Lynch syndrome, (2) have colon cancer, (3) have endometrial cancer meeting clinical criteria for Lynch syndrome, (4) are at-risk relatives of patients with Lynch syndrome, (5) are without colon cancer but with a family history meeting Amsterdam or Revised Bethesda criteria, or documentation of 5% or higher predicted risk of the syndrome on a validated risk prediction model, who receive genetic testing for MMR genes, the evidence includes an Agency for Healthcare Research and Quality report, a supplemental assessment to that report by the Evaluation of Genomic Applications in Practice and Prevention Working Group, and an Evaluation of Genomic Applications in Practice and Prevention recommendation for genetic testing in colorectal cancer (CRC). Relevant outcomes are OS, disease-specific survival, and test accuracy and validity. A chain of evidence from well-designed experimental nonrandomized studies is adequate to demonstrate the clinical utility of testing unaffected (without cancer) first- and second-degree relatives of patients with Lynch syndrome who have a known variant in an MMR gene, in that counseling has been shown to influence testing and surveillance choices among unaffected family members of Lynch syndrome patients. One long-term, nonrandomized controlled study and a cohort study of Lynch syndrome family members found significant reductions in CRC among those who followed recommended colonic surveillance. A positive genetic test for an MMR variant can also lead to changes in the management of other Lynch syndrome malignancies. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who warrant Lynch testing, screen negative on MMR testing, but positive for microsatellite instability (MSI) and lack MSH2 protein expression who receive genetic testing for *EPCAM* variants, the evidence includes variant prevalence studies and case series. Relevant outcomes are OS, disease-specific survival, and test accuracy and validity. Studies have shown an association between *EPCAM* variants and Lynch-like disease in families, and the cumulative risk for CRC is similar to carriers of an *MSH2* variant. Identification of an *EPCAM* variant could lead to changes in management that improve health outcomes. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have CRC in whom MLH1 protein is not expressed on immunohistochemical (IHC) analysis and who receive genetic testing for *BRAF* V600E or *MLH1* promoter methylation, the evidence includes case series. Relevant outcomes are OS, disease-specific survival, and test accuracy and validity. Studies have shown, with high sensitivity and specificity, an

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association between *BRAF* V600E variant and *MLH1* promoter methylation with sporadic CRC. Therefore, this type of testing could eliminate the need for further genetic testing or counseling for Lynch syndrome. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who (1) are suspected of JPS or PJS or (2) are at-risk relatives of patients suspected of or diagnosed with juvenile polyposis syndrome (JPS) or Peutz-Jeghers syndrome (PJS) who receive genetic testing for *SMAD4*, *BMPR1A*, or *STK11* genes, respectively, the evidence includes multiple observational studies. Relevant outcomes are OS, disease-specific survival, and test accuracy and validity. Studies have shown, with high sensitivity and specificity, an association between *SMAD4* and *BMPR1A* and *STK11* variants with JPS and PJS, respectively. Direct evidence of clinical utility for genetic testing of JPS or PJS is not available. Genetic testing may have clinical utility by avoiding burdensome and invasive endoscopic examinations, release from intensified screening programs resulting in psychological relief, and improving health outcomes by identifying currently unaffected atrisk family members who require intense surveillance or prophylactic colectomy. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

JAK2, MPL, and CALR Testing for Myeloproliferative Neoplasms

For individuals with a suspected myeloproliferative neoplasm (MPN) who receive genetic testing for *JAK2*, the evidence includes case series, retrospective studies, meta-analyses, and randomized controlled trials. Relevant outcomes are overall survival (OS), disease-specific survival, test accuracy and validity, and resource utilization. For patients with suspected Phnegative MPN, *JAK2* variants are found in nearly 100% of those with polycythemia vera (PV), 60% to 65% of those with essential thrombocythemia (ET), and 60% to 65% of those with primary myelofibrosis (PMF). In individuals with suspected MPN, a positive genetic test for *JAK2* satisfies a major criterion for the International Consensus Classification (2022) and World Health Organization (WHO) 2022 (5th edition) classification for Ph-negative MPNs and eliminates secondary or reactive causes of erythrocytosis and thrombocythemia from the differential diagnosis. The presence of a documented *JAK2* variant may aid in the selection of ruxolitinib, a *JAK2* inhibitor; ruxolitinib, however, is classified as second-line therapy. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals with a suspected MPN who receive genetic testing for MPL, the evidence includes case series and retrospective studies. Relevant outcomes are OS, disease-specific survival, test accuracy and validity, and resource utilization. For patients with suspected Phnegative MPN, MPL variants are found in approximately 5% of those with ET and PMF. In individuals with suspected MPN, a positive genetic test for MPL satisfies a major criterion for the International Consensus Classification (2022) and WHO (2022, 5th edition) classification for ET and PMF and eliminates secondary or reactive causes of thrombocythemia from the differential diagnosis. The goal of ET treatment is to alleviate symptoms and minimize thrombotic events and bleeding irrespective of MPL variant status. For PMF, hematopoietic cell transplantation is the only treatment with curative potential while most other treatment options focus on symptom alleviation. However, in both ET and PMF, establishing the diagnosis through MPL genetic testing does not in and of itself result in changes in

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management that would be expected to improve the net health outcome. Thus, the clinical utility has not been established. The evidence is insufficient to determine that the technology results in an improvement in the net health outcome.

For individuals with a suspected MPN who receive genetic testing for *CALR*, the evidence includes case series and retrospective studies. Relevant outcomes are OS, disease-specific survival, test accuracy and validity, and resource utilization. For patients with suspected Phnegative MPN, *CALR* variants are found in approximately 20% to 25% of those with ET and PMF. For individuals with suspected MPN, a positive genetic test for *CALR* satisfies a major criterion for the International Consensus Classification (2022) and WHO (2022, 5th edition) classification for ET and PMF and eliminates secondary or reactive causes of thrombocythemia from the differential diagnosis. The goal of ET treatment is to alleviate symptoms and minimize thrombotic events and bleeding irrespective of *CALR* variant status. For PMF, hematopoietic cell transplantation is the only treatment with curative potential while most other treatment options focus on symptom alleviation. However, in both ET and PMF, establishing the diagnosis through *CALR* genetic testing does not result in changes in management that would be expected to improve the net health outcome. Thus, the clinical utility has not been established. The evidence is insufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have suspected chronic myelogenous leukemia (CML) who receive *BCR-ABL1* fusion gene qualitative testing to confirm the diagnosis and establish a baseline for monitoring treatment, the evidence includes validation studies. Relevant outcome is test validity. The sensitivity of testing with reverse transcription-polymerase chain reaction is high compared with conventional cytogenetics. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have a diagnosis of CML who receive *BCR-ABL1* fusion gene quantitative testing at appropriate intervals for monitoring treatment response and remission, the evidence includes a systematic review and nonrandomized trials. Relevant outcomes are disease-specific survival, test validity, and change in disease status. Studies have shown high sensitivity of this type of testing and a strong correlation with outcomes, including the risk of disease progression and survival, which may stratify patients to different options for disease management. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

BCR-ABL1 Testing in Chronic Myelogenous Leukemia and Acute Lymphoblastic Leukemia

For individuals who have a diagnosis of CML with an inadequate initial response, loss of response, and/or disease progression who receive an evaluation for *ABL* kinase domain (KD) single nucleotide variants to assess for tyrosine kinase inhibitor (TKI) resistance, the evidence includes a systematic review and retrospective cohort study. Relevant outcomes are disease-specific survival, test validity, and medication use. The systematic review and case series evaluated pharmacogenetics testing for TKIs and reported the presence of KD single nucleotide variants detected at imatinib failure. These studies have shown a correlation between certain types of variants, treatment response, and the selection of subsequent

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treatment options. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have a diagnosis of Philadelphia chromosome (Ph)-positive acute lymphoblastic leukemia (ALL) who receive *BCR-ABL1* fusion gene quantitative testing at baseline before and during treatment to monitor treatment response and remission, the evidence includes prospective and retrospective cohort studies and case series. Relevant outcomes are disease-specific survival, test validity, and change in disease status. As with CML, studies have shown high sensitivity for this type of testing and a strong correlation with outcomes, including the risk of disease progression, which may stratify patients to different treatment options. Also, evidence of treatment resistance or disease recurrence directs a change in medication. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who have Ph-positive ALL and signs of treatment failure or disease progression who receive an evaluation for *ABL1* KD single nucleotide variants to assess for TKI resistance, the evidence includes case series. Relevant outcomes are test validity and medication use. Studies have shown that specific imatinib-resistant variants are insensitive to 1 or more of the second-generation TKIs; these variants are used to guide medication selection. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

Carrier Screening for Genetic Diseases

For individuals who are asymptomatic but at risk for having offspring with an inherited X-linked or autosomal recessive genetic disorder who receive targeted risk-based carrier screening, the evidence includes studies supporting clinical validity and clinical utility. Relevant outcomes are test validity and changes in reproductive decision making. Results of carrier testing can be used to inform reproductive decisions such as preimplantation genetic diagnosis, in vitro fertilization, not having a child, invasive prenatal testing, adoption, or pregnancy termination. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

For individuals who are either at increased risk or population risk for having offspring with an inherited X-linked or autosomal recessive genetic disorder who receive a non-targeted carrier screening panel, the evidence includes studies supporting clinical validity and clinical utility. Relevant outcomes are test validity and changes in reproductive decision making. Studies have found that non-targeted carrier screening identifies more carriers and more potentially affected fetuses. Many of the genes in carrier screening panels do not meet the American College of Obstetricians and Gynecologists (ACOG) consensus-driven criteria of at least a 1% carrier rate for all ethnic groups. However, non-targeted testing can address the discrepancies between self-reported ethnicity and genetic ancestry in an ethnically mixed population. As panels become larger the likelihood of being identified as a carrier of a rare genetic disorder increases, leading to an at-risk couple rate of nearly 2% for having an offspring with a recessive or X-linked disorder. Many, though notably not all, of these rare genetic disorders are associated with severe or profound symptoms including shortened lifespan and intellectual or physical disability. With adequate genetic counseling, carrier screening panels can inform reproductive choices, and observational studies have shown that

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a majority of couples would consider intervention that depends on the severity of the condition. Therefore, non-targeted carrier screening panels for severe recessive and X-linked genetic disorders can have a significant clinical impact. The evidence is sufficient to determine that the technology results in an improvement in the net health outcome.

Policy Guidelines

Germline Genetic Testing for Hereditary Breast/Ovarian Cancer Syndrome and Other High-Risk Cancers (BRCA1, BRCA2, PALB2)

Close Relatives

Close relatives are blood related family members including 1st-, 2nd-, and 3rd-degree relatives on the same side of the family (maternal or paternal).

- 1st-degree relatives are parents, siblings, and children.
- 2nd-degree relatives are grandparents, aunts, uncles, nieces, nephews, grandchildren, and half-siblings.
- 3rd-degree relatives are great-grandparents, great-aunts, great-uncles, great-grandchildren, and first cousins.

Prostate Cancer Risk Groups

Risk groups for prostate cancer in this policy include high-risk groups and very-high-risk groups.

- High-risk group: no very-high-risk features and are T3a (American Joint Committee on Cancer staging T3a = tumor has extended outside of the prostate but has not spread to the seminal vesicles); OR Grade Group 4 or 5; OR prostate specific antigen of 20 ng/mL or greater.
- Very-high-risk group: T3b-T4 (tumor invades seminal vesicle(s); or tumor is fixed or invades adjacent structures other than seminal vesicles such as external sphincter, rectum, bladder, levator muscles, and/or pelvic wall); OR Primary Gleason Pattern 5; OR 2 or 3 high-risk features; OR greater than 4 cores with Grade Group 4 or 5.

Recommended Testing Strategies

Individuals who meet criteria for genetic testing as outlined in the policy statements above should be tested for variants in *BRCA1*, *BRCA2*, and *PALB2*. Recommended strategies are listed below.

- In individuals with a known familial *BRCA* or *PALB2* variant, targeted testing for the specific variant is recommended.
- In individuals with unknown familial BRCA or PALB2 variant:
 - To identify clinically significant variants, NCCN advises testing a relative who has early-onset disease, bilateral disease, or multiple primaries, because that individual has the highest likelihood of obtaining a positive test result. Unless the affected individual is a member of an ethnic group for which particular founder pathogenic or likely pathogenic variants are known, comprehensive

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- genetic testing (ie, full sequencing of the genes and detection of large gene rearrangements) should be performed.
- If no living family member with breast or ovarian cancer exists, NCCN suggests testing first- or second-degree family members affected with cancer thought to be related to deleterious BRCA1 or BRCA2 variants (eg, prostate cancer, pancreatic cancer, melanoma).
- If no familial variant can be identified, 2 possible testing strategies are:
 - Full sequencing of BRCA1 and BRCA2 followed by testing for large genomic rearrangements (deletions, duplications) only if sequencing detects no variant (negative result). More than 90% of BRCA variants will be detected by full sequencing.
 - Alternatively, simultaneous full sequencing and testing for large genomic rearrangements (also known as comprehensive BRCA testing; see Comprehensive Variant Analysis below) may be performed as is recommended by NCCN. Comprehensive testing can detect 92.5% of BRCA1 or BRCA2 variants.
- Testing for BRCA1, BRCA2, and PALB2 through panel testing over serial testing might
 be preferred for efficiency. Multi-gene panels often include genes of moderate or low
 penetrance, and genes with limited evidence on which to base management decisions.
 When considering a gene panel, NCCN recommends use of "tailored panels that are
 disease-focused and include clinically actionable cancer susceptibility genes".
- Ashkenazi Jewish descent
 - In individuals of known Ashkenazi Jewish descent, one approach is to test for the 3 known founder mutations (185delAG and 5182insC in *BRCA1*; 6174delT in *BRCA2*) first, if testing is negative for founder mutations and if the individual's ancestry also includes non-Ashkenazi ethnicity (or if other *BRCA1*/2 testing criteria are met), comprehensive genetic testing should be considered.
- Testing strategy may also include testing individuals not meeting the above criteria
 who are adopted and have limited medical information on biological family members,
 individuals with small family structure, and individuals with presumed paternal
 transmission.

High-Risk Ethnic Groups

Testing of eligible individuals who belong to ethnic populations in which there are well-characterized founder mutations should begin with tests specifically for these variants. For example, founder mutations account for approximately three-quarters of the *BRCA* variants found in Ashkenazi Jewish populations. When testing for founder mutations is negative, a comprehensive variant analysis should then be performed.

Testing Unaffected Individuals

In unaffected family members of potential *BRCA* or *PALB2* variant families, most test results will be negative and uninformative. Therefore, it is strongly recommended that an *affected* family member be tested first whenever possible to adequately interpret the test. Should a *BRCA* or *PALB2* variant be found in an affected family member(s), DNA from an *unaffected* family member can be tested specifically for the same variant of the affected family member without having to sequence the entire gene. Interpreting test results for an

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unaffected family member without knowing the genetic status of the family may be possible in the case of a positive result for an established disease-associated variant but leads to difficulties in interpreting negative test results (uninformative negative) or variants of uncertain significance because the possibility of a causative *BRCA* or *PALB2* variant is not ruled out.

Testing Minors

The use of genetic testing for *BRCA1*, *BRCA2*, or *PALB2* variants for identifying hereditary breast and ovarian cancer syndrome has limited or no clinical utility in minors, because there is no change in management for minors as a result of knowledge of the presence or absence of a deleterious variant. In addition, there are potential harms related to stigmatization and discrimination. See policy 2.04.128 regarding testing of *BRCA1*, *BRCA2*, and *PALB2* for Fanconi anemia. See policies 2.04.148, 2.04.151, 2.04.155, and 2.04.156 regarding genetic testing to guide targeted therapy.

Prostate Cancer

Individuals with *BRCA* or *PALB2* variants have an increased risk of prostate cancer, and individuals with known *BRCA* or *PALB2* variants may, therefore, consider more aggressive screening approaches for prostate cancer.

Genetic Testing for Lynch Syndrome and Other Inherited Colon Cancer Syndromes

Testing At-Risk Relatives

Due to the high lifetime risk of cancer of most genetic syndromes discussed in this policy, "atrisk relatives" primarily refers to first-degree relatives. However, some judgment must be permitted, eg, in the case of a small family pedigree, when extended family members may need to be included in the testing strategy. Family history might include at least 2 second-degree relatives with a Lynch syndrome-related cancer, including at least 1 diagnosed before 50 years of age, or at least 3 second-degree relatives with a Lynch syndrome-related cancer, regardless of age.

Targeted Familial Variant Testing

It is recommended that, when possible, initial genetic testing for familial adenomatous polyposis (FAP) or Lynch syndrome be performed in an affected family member, so that testing in unaffected family members can focus on the variant found in the affected family member (see Benefit Application section). If an affected family member is not available for testing, testing should begin with an unaffected family member most closely related to an affected family member.

In many cases, genetic testing for *MUTYH* gene variants should first target the specific variants *Y165C* and *G382D*, which account for more than 80% of variants in white populations, and subsequently, proceed to sequence only as necessary. However, in other ethnic populations, proceeding directly to sequencing is appropriate.

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Evaluation for Lynch Syndrome

For patients with colorectal cancer (CRC) or endometrial cancer being evaluated for Lynch syndrome, the microsatellite instability (MSI) test or the immunohistochemical (IHC) test with or without *BRAF* gene variant testing, or methylation testing, should be used as an initial evaluation of tumor tissue before mismatch repair (MMR) gene analysis. Both tests are not necessary. Proceeding to MMR gene sequencing would depend on the results of MSI or IHC testing. In particular, IHC testing may help direct which MMR gene likely contains a variant, if any, and may also provide additional information if MMR genetic testing is inconclusive. For further information on tumor tissue test results, interpretation, and additional testing options, see the NCCN [National Comprehensive Cancer Network] clinical care guidelines on genetic/familial high-risk assessment: colorectal.

When indicated, genetic sequencing for MMR gene variants should begin with MLH1 and MSH2 genes, unless otherwise directed by the results of IHC testing. Standard sequencing methods will not detect large deletions or duplications; when MMR gene variants are expected based on IHC or MSI studies, but none are found by standard sequencing, additional testing for large deletions or duplications is appropriate.

The Amsterdam II Clinical Criteria (all criteria must be fulfilled) are the most stringent for defining families at high risk for Lynch syndrome:

- 3 or more relatives with an associated cancer (CRC, or cancer of the endometrium, small intestine, ureter, or renal pelvis);
- 1 should be a first-degree relative of the other 2;
- 2 or more successive generations affected;
- 1 or more relatives diagnosed before the age of 50 years;
- FAP should be excluded in cases of CRC;
- Tumors should be verified by pathologic examination.
- Modifications:
 - EITHER: very small families, which cannot be further expanded, can be considered to have hereditary nonpolyposis colorectal cancer (HNPCC) with only 2 CRCs in first-degree relatives if at least 2 generations have the cancer and at least 1 case of CRC was diagnosed by the age of 55 years;
 - OR: in families with 2 first-degree relatives affected by CRC, the presence of a third relative with an unusual early-onset neoplasm or endometrial cancer is sufficient.

The Revised Bethesda Guidelines (fulfillment of any criterion meets guidelines) are less stringent than the Amsterdam criteria and are intended to increase the sensitivity of identifying at-risk families. The Bethesda guidelines are also considered more useful in identifying which patients with CRC should have their tumors tested for MSI and/or IHC:

- CRC diagnosed in a patient who is younger than 50 years old;
- Presence of synchronous or metachronous CRC or other HNPCC-associated tumors, a regardless of age;

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- CRC with high MSI histology diagnosed in a patient younger than 60 years old;
- CRC diagnosed in 1 or more first-degree relatives with a Lynch syndrome-associated tumor, with 1 of the cancers being diagnosed before 50 years of age;
- CRC diagnosed in 2 or more first or second-degree relatives with HNPCC-related tumors, a regardless of age.

^a HNPCC-related tumors include colorectal, endometrial, stomach, ovarian, pancreas, ureter and renal pelvis, biliary tract, brain (usually glioblastoma as seen in Turcot syndrome), sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome, and carcinoma of the small bowel.

Multiple risk prediction models that provide quantitative estimates of the likelihood of an MMR variant are available such MMRpro, PREMM5, or MMRpredict. National Comprehensive Cancer Network guidelines recommend (category 2A) testing for Lynch syndrome in individuals with a 5% or higher predicted risk of the syndrome on these risk prediction models.

Genetic Counseling

Genetic counseling is primarily aimed at patients who are at risk for inherited disorders, and experts recommend formal genetic counseling in most cases when genetic testing for an inherited condition is considered. The interpretation of the results of genetic tests and the understanding of risk factors can be very difficult and complex. Therefore, genetic counseling will assist individuals in understanding the possible benefits and harms of genetic testing, including the possible impact of the information on the individual's family. Genetic counseling may alter the utilization of genetic testing substantially and may reduce inappropriate testing. Genetic counseling should be performed by an individual with experience and expertise in genetic medicine and genetic testing methods.

JAK2, MPL, and CALR Testing for Myeloproliferative Neoplasms

Testing Strategy

Individuals suspected to have polycythemia vera (PV) should first be tested for the most common finding, *JAK2* V617F. If the testing is negative, further testing to detect other *JAK2* tyrosine kinase variants (eg, in exon 12) is warranted.

Individuals suspected to have essential thrombocythemia or primary myelofibrosis should first be tested for *JAK2* variants, as noted. If testing is negative, further testing to detect *MPL* and *CALR* variants is warranted.

Criteria for Polycythemia Vera Testing

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Based on the World Health Organization (WHO) and International Consensus Classification major and minor criteria (see Table 1), documentation of serum erythropoietin level below the reference range for normal meets a minor criterion for PV. Therefore, serum erythropoietin testing is recommended before *JAK2* testing.

Table 1. World Health Organization 5th Edition and the International Consensus Classification Diagnostic Criteria for Polycythemia Vera

Major Criteria

- Increased hemoglobin level (>16.5 g/dL in men or >16.0 g/dL in women); or
- Increased hematocrit (>49% in men or >48% in women); or
- Bone marrow biopsy showing hypercellularity for age with trilineage maturation, including prominent erythroid, granulocytic, and megakaryocytic proliferation with pleomorphic, mature megakaryocytes (differences in size)
- JAK2 V617F or JAK2 exon 12 variant detected

Minor Criterion

• Serum erythropoietin level below the reference range for normal

BCR-ABL1 Testing in Chronic Myelogenous Leukemia and Acute Lymphoblastic Leukemia

Diagnosis of Chronic Myelogenous Leukemia and Acute Lymphoblastic Leukemia Qualitative molecular confirmation of the cytogenetic diagnosis (ie, detection of the Philadelphia chromosome) is necessary for accurate diagnosis of chronic myelogenous leukemia (CML). Identification of the Philadelphia chromosome is not necessary to diagnose acute lymphoblastic leukemia (ALL); however, molecular phenotyping is usually performed at the initial assessment (see Determining Baseline RNA Transcript Levels and Subsequent Monitoring subsection).

Distinction between molecular variants (ie, p190 vs p210) is necessary for accurate results in subsequent monitoring assays.

Determining Baseline RNA Transcript Levels and Subsequent Monitoring

Determination of *BCR-ABL1* messenger RNA transcript levels should be done by quantitative real-time reverse transcription-polymerase chain reaction-based assays and reported results should be standardized according to the International Scale.

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For CML, testing is appropriate at baseline before the start of imatinib treatment, and testing is appropriate every 3 months when the individual is responding to treatment. After a complete cytogenetic response is achieved, testing is recommended every 3 months for 2 years, then every 3 to 6 months thereafter during treatment.

Without a complete cytogenetic response, continued monitoring at 3-month intervals during treatment is recommended. It has been assumed that the same time points for monitoring imatinib are appropriate for dasatinib and nilotinib and will likely also be applied to bosutinib and ponatinib.

More frequent monitoring is indicated for individuals diagnosed with CML who are in complete molecular remission and are not undergoing treatment with a tyrosine kinase inhibitor (TKI). For ALL, the optimal timing remains unclear and depends on the chemotherapy regimen used.

Tyrosine Kinase Inhibitor Resistance

For CML, inadequate initial response to TKIs is defined as failure to achieve a complete hematologic response at 3 months, only minor cytogenetic response at 6 months, or major (rather than complete) cytogenetic response at 12 months.

Unlike in CML, ALL resistance to TKIs is less well studied. In individuals with ALL receiving a TKI, a rise in the *BCR-ABL* mRNA level while in hematologic complete response or clinical relapse warrants variant analysis.

Loss of response to TKIs is defined as hematologic relapse, cytogenetic relapse, or 1-log increase in *BCR-ABL1* transcript ratio and therefore loss of major molecular response.

Kinase domain single nucleotide variant testing is usually offered as a single test to identify T315I variant or as a panel (that includes T315I) of the most common and clinically important variants.

Carrier Screening for Genetic Diseases

Carrier screening (targeted or non-targeted) is only medically necessary once per lifetime. Exceptions may be considered if advances in technology support medical necessity for retesting.

Targeted carrier screening for autosomal recessive or X-linked conditions is also called risk-based test or ethnic-based testing. If targeted testing is performed by a panel, the most appropriate panel code available should be used. The panel and the panel billing code should include *CFTR* and *SMN1*.

Non-targeted carrier screening applies to persons of any risk including average risk. Any panel using 81443 for non-targeted carrier screening must include the *CFTR* and *SMN1* genes. Non-targeted carrier screening panels should include the minimum number of genes but not exceed the maximum number of genes recommended by professional guidelines from the American College of Obstetricians and Gynecologists (ACOG; 2-22 conditions) or the American College of Medical Genetics and Genomics (ACMG; 113 genes).

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The Committee (reaffirmed in 2023) states that "Ethnic-specific, panethnic, and expanded carrier screening are acceptable strategies for prepregnancy and prenatal carrier screening" and offered the following summary pertaining to expanded carrier screening: "Given the multitude of conditions that can be included in expanded carrier screening panels, the disorders selected for inclusion should meet several of the following consensus-determined criteria: have a carrier frequency of 1 in 100 or greater, have a well-defined phenotype, have a detrimental effect on quality of life, cause cognitive or physical impairment, require surgical or medical intervention, or have an onset early in life. Additionally, screened conditions should be able to be diagnosed prenatally and may afford opportunities for antenatal intervention to improve perinatal outcomes, changes to delivery management to optimize newborn and infant outcomes, and education of the parents about special care needs after birth. Carrier screening panels should not include conditions primarily associated with a disease of adult onset."[ACOG Committee Opinion No. 690; PMID: 28225425]

The ACOG guideline includes a list of 22 conditions deemed reasonable to include in a carrier screening panel (see <u>Table 2</u>). While there is no agreed upon definition of severity across professional societies, these 22 conditions have severity that would be deemed profound or severe per publication based on previous work by ACMG and cited by the most recent ACMG guidelines.[Lazarin et al (2014); PMID: 25494330][Gregg et al (2021); PMID 34285390] All but one condition deemed reasonable by ACOG (alpha-thalassemia) would be classified as profound or severe based on collaborative clinical expert application of a trait-based algorithm; however, in this work it is not clear if the alpha-thalassemia genes *HBA1/HBA2* were classified based on hemoglobin Bart hydrops fetalis syndrome or hemoglobin H disease.[Arjunan et al (2020); PMID: 32474937] Carrier testing of autosomal recessive genes associated with severe disease with carrier frequency of greater than 1/100 is estimated to identify 82% of at-risk couples.[Guo et al (2019); PMID: 30846881]

In 2021, the ACMG recommended that the phrase "expanded carrier screening" be replaced by "carrier screening" as expanded carrier screening is not well or precisely defined by professional organizations. [Gregg et al (2021); PMID 34285390] Previously, ACMG has defined expanded panels as those that use next-generation sequencing to screen for variants in many genes, as opposed to gene-by-gene screening (eg, ethnic-specific screening or panethnic testing for cystic fibrosis).

The updated ACMG guideline now recommends a multi-tier approach to carrier screening for autosomal recessive and X-linked conditions, incorporating recommendations from the ACOG Committee Opinion 691 (2017; reaffirmed in 2023),[ACOG Committee Opinion No. 691; PMID: 28225426] to enhance communication and precision while advancing equity in carrier screening (see Table PG1).[Gregg et al (2021); PMID 34285390] The consensus group recognized no accepted standard in defining the severity of various conditions; and, based on previously published work, use the following definitions: (1) profound: shortened lifespan during infancy or childhood, intellectual disability; (2) severe: death in early adulthood, impaired mobility or a [disabling] malformation involving an internal organ; (3) moderate: neurosensory impairment, immune deficiency or cancer, mental illness, dysmorphic features; and (4) mild: not meeting one of those described.[Lazarin et al (2014); PMID: 25494330]

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The ACMG consensus group recommends offering Tier 3 carrier screening ($\geq 1/200$ carrier frequency + Tier 2; see Table PG1) to all pregnant patients and those planning a pregnancy. Carrier testing of autosomal recessive genes associated with severe disease with carrier frequency greater than 1/100 is estimated to identify 82% of at-risk couples, and identify 93% of at-risk couples when testing for genes with greater than 1/200 carrier frequency.[Guo et al (2019); PMID: 30846881] The ACMG Tier 3 recommendations were based on estimates that moving from Tier 2 ($\geq 1/100$ carrier frequency) to Tier 3 (1/200 carrier frequency) provided additional identification of 4-9/10,000 at-risk couples depending on the endogamous population examined. When the population evaluated was weighted by U.S. census data, at-risk couples identified increased by 6 per 10,000 couples when moving from the Tier 2 ($\geq 1/100$) carrier frequency to that of Tier 3 ($\geq 1/200$). Assuming ~4 million births per year, this translates to an annual increase of identifying 2,400 additional U.S. couples.

The ACMG consensus group specified gene recommendations which include testing for 97 autosomal recessive genes and 16 X-linked genes, all of which associate with disorders of moderate, severe, or profound severity and are of 1/200 or greater carrier frequency. Nontargeted carrier screening panels that test for genes beyond this provide diminishingly small results, and pleiotropy, locus heterogeneity, variant interpretation, and poor genotype-phenotype correlation may disproportionately impact the ability to provide accurate prognostic information. [Gregg et al (2021); PMID 34285390]

Additionally, the recommendations include that male partners of pregnant women and those planning a pregnancy may be offered Tier 3 carrier screening for autosomal recessive conditions when carrier screening is performed simultaneously with their female partner. Tier 4 screening may be offered when a pregnancy stems from a known or possible consanguineous relationship (second cousins or closer) or when family or personal medical history warrants. The ACMG does not recommend offering Tier 1 and/or Tier 2 screening, because these do not provide equitable evaluation of all racial/ethnic groups, or the routine offering of Tier 4 panels.

Testing Strategy

After testing the proband, targeted testing on the reproductive partner is preferred. Testing only applies to genes meeting criteria outlined above. If a lab does a more extensive test, then testing for other findings in the reproductive partner would not meet criteria. In general, carrier screening can be done once per lifetime. However, if only targeted or limited testing was done previously, then a more general non-targeted panel could be performed, particularly in cases where there is a new reproductive partner. In this case it is likely that genes could be re-tested.

Genetic Counseling

Genetic counseling is primarily aimed at patients who are at risk for inherited disorders, and experts recommend formal genetic counseling in most cases when genetic testing for an inherited condition is considered. The interpretation of the results of genetic tests and the understanding of risk factors can be very difficult and complex. Therefore, genetic counseling will assist individuals in understanding the possible benefits and harms of genetic testing, including the possible impact of the information on the individual's family. Genetic counseling may alter the utilization of genetic testing substantially and may reduce inappropriate

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testing. Genetic counseling should be performed by an individual with experience and expertise in genetic medicine and genetic testing methods. Carrier screening with appropriate genetic counseling is performed in adults.

Table 2. Example of an Expanded Carrier Screening Panel

Condition	Carrier Frequency in General Population	Carrier Frequency in Specific Ethnic Groups
α-thalassemia	Unknown	African (particularly sub-Saharan): 1 in 3 Mediterranean: 1 in 30 Southeast Asian and Middle Eastern: 1 in 20
ß-thalassemia	Unknown	African American: <1 in 8 Ashkenazi Jewish: Varied Asian: 1 in 20 Mediterranean: 1 in 7
Bloom syndrome	<1 in 500	Ashkenazi Jewish: 1 in 100
Canavan disease	<1 in 150	Ashkenazi Jewish: 1 in 41
Cystic fibrosis	Unknown	African American: 1 in 61 Asian: 1 in 94 Ashkenazi Jewish: 1 in 24 Caucasian: 1 in 25 Hispanic: 1 in 58
Familial dysautonomia	<1 in 500	Ashkenazi Jewish: 1 in 31
Familial hyperinsulinism	<1 in 150	Ashkenazi Jewish: 1 in 52
Fanconi anemia C	<1 in 790	Ashkenazi Jewish: 1 in 89
Fragile X syndrome	1 in 259	
Galactosemia	1 in 87	Ashkenazi Jewish: 1 in 127
Gaucher disease	<1 in 100	Ashkenazi Jewish: 1 in 15
Glycogen storage disease type 1A	<1 in 150	Ashkenazi Jewish: 1 in 71
Joubert syndrome	<1 in 500	Ashkenazi Jewish: 1 in 92
Medium-chain acyl-CoA dehydrogenase deficiency	Unknown	Caucasian: 1 in 50
Maple syrup urine disease types 1A and 1B	1 in 240	Ashkenazi Jewish: 1 in 81 (type 1B) Mennonite: 1 in 10 (type 1A- BCKDHA p.Y438N)
Mucolipidosis IV	<1 in 500	Ashkenazi Jewish: 1 in 96
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Niemann-Pick disease type A	<1 in 500	Ashkenazi Jewish: 1 in 90
Phenylketonuria	Unknown	Caucasian: 1 in 50 Irish: 1 in 34
Sickle cell anemia	Unknown	African American: 1 in 10
Smith-Lemli-Opitz syndrome	Unknown	Caucasian: 1 in 70
Spinal muscular atrophy	Unknown	African American: 1 in 66 Asian: 1 in 53 Ashkenazi Jewish: 1 in 41 Caucasian: 1 in 35 Hispanic: 1 in 117
Tay-Sachs disease	1 in 300	Ashkenazi Jewish: 1 in 30 French Canadian and Cajun: 1 in 30

References

Germline Genetic Testing for Hereditary Breast/Ovarian Cancer Syndrome and Other High-Risk Cancers (BRCA1, BRCA2, PALB2)

- 1. Chapman-Davis E, Zhou ZN, Fields JC, et al. Racial and Ethnic Disparities in Genetic Testing at a Hereditary Breast and Ovarian Cancer Center. J Gen Intern Med. Jan 2021; 36(1): 35-42. PMID 32720237
- 2. Winchester DP. Breast cancer in young women. Surg Clin North Am. Apr 1996; 76(2): 279-87. PMID 8610264
- 3. Frank TS, Deffenbaugh AM, Reid JE, et al. Clinical characteristics of individuals with germline mutations in BRCA1 and BRCA2: analysis of 10,000 individuals. J Clin Oncol. Mar 15 2002; 20(6): 1480-90. PMID 11896095
- 4. Langston AA, Malone KE, Thompson JD, et al. BRCA1 mutations in a population-based sample of young women with breast cancer. N Engl J Med. Jan 18 1996; 334(3): 137-42. PMID 8531967
- 5. Malone KE, Daling JR, Thompson JD, et al. BRCA1 mutations and breast cancer in the general population: analyses in women before age 35 years and in women before age 45 years with first-degree family history. JAMA. Mar 25 1998; 279(12): 922-9. PMID 9544766
- 6. Ford D, Easton DF, Stratton M, et al. Genetic heterogeneity and penetrance analysis of the BRCA1 and BRCA2 genes in breast cancer families. The Breast Cancer Linkage Consortium. Am J Hum Genet. Mar 1998; 62(3): 676-89. PMID 9497246
- 7. Gershoni-Baruch R, Patael Y, Dagan A, et al. Association of the I1307K APC mutation with hereditary and sporadic breast/ovarian cancer: more questions than answers. Br J Cancer. Jul 2000; 83(2): 153-5. PMID 10901363
- 8. Warner E, Foulkes W, Goodwin P, et al. Prevalence and penetrance of BRCA1 and BRCA2 gene mutations in unselected Ashkenazi Jewish women with breast cancer. J Natl Cancer Inst. Jul 21 1999; 91(14): 1241-7. PMID 10413426

Page 25 of 55

- 9. Hartge P, Struewing JP, Wacholder S, et al. The prevalence of common BRCA1 and BRCA2 mutations among Ashkenazi Jews. Am J Hum Genet. Apr 1999; 64(4): 963-70. PMID 10090881
- 10. Hodgson SV, Heap E, Cameron J, et al. Risk factors for detecting germline BRCA1 and BRCA2 founder mutations in Ashkenazi Jewish women with breast or ovarian cancer. J Med Genet. May 1999; 36(5): 369-73. PMID 10353781
- 11. Moslehi R, Chu W, Karlan B, et al. BRCA1 and BRCA2 mutation analysis of 208 Ashkenazi Jewish women with ovarian cancer. Am J Hum Genet. Apr 2000;66(4):1259-1272. PM
- 12. de Ruijter TC, Veeck J, de Hoon JP, et al. Characteristics of triple-negative breast cancer. J Cancer Res Clin Oncol. Feb 2011; 137(2): 183-92. PMID 21069385
- 13. Kandel MJ, Stadler D, Masciari S, et al. Prevalence of BRCA1 mutations in triple negative breast cancer (BC) [abstract 508]. J Clin Oncol. 2006;24(18S):508.
- 14. Young SR, Pilarski RT, Donenberg T, et al. The prevalence of BRCA1 mutations among young women with triple-negative breast cancer. BMC Cancer. Mar 19 2009; 9: 86. PMID 19298662
- 15. Gonzalez-Angulo AM, Timms KM, Liu S, et al. Incidence and outcome of BRCA mutations in unselected patients with triple receptor-negative breast cancer. Clin Cancer Res. Mar 01 2011; 17(5): 1082-9. PMID 21233401
- 16. Xia B, Sheng Q, Nakanishi K, et al. Control of BRCA2 cellular and clinical functions by a nuclear partner, PALB2. Mol Cell. Jun 23 2006; 22(6): 719-729. PMID 16793542
- 17. Antoniou AC, Casadei S, Heikkinen T, et al. Breast-cancer risk in families with mutations in PALB2. N Engl J Med. Aug 07 2014; 371(6): 497-506. PMID 25099575
- 18. Catucci I, Peterlongo P, Ciceri S, et al. PALB2 sequencing in Italian familial breast cancer cases reveals a high-risk mutation recurrent in the province of Bergamo. Genet Med. Sep 2014; 16(9): 688-94. PMID 24556926
- 19. Casadei S, Norquist BM, Walsh T, et al. Contribution of inherited mutations in the BRCA2-interacting protein PALB2 to familial breast cancer. Cancer Res. Mar 15 2011; 71(6): 2222-9. PMID 21285249
- 20. Cybulski C, Kluźniak W, Huzarski T, et al. Clinical outcomes in women with breast cancer and a PALB2 mutation: a prospective cohort analysis. Lancet Oncol. Jun 2015; 16(6): 638-44. PMID 25959805
- 21. Blue Cross and Blue Shield Association Technology Evaluation Center (TEC). BRCA1 and BRCA2 testing to determine the risk of breast and ovarian cancer. TEC Assessments. 1997; Volume 12:Tab 4.
- 22. Zhu Y, Wu J, Zhang C, et al. BRCA mutations and survival in breast cancer: an updated systematic review and meta-analysis. Oncotarget. Oct 25 2016; 7(43): 70113-70127. PMID 27659521
- 23. Nelson HD, Fu R, Goddard K, et al. Risk Assessment, Genetic Counseling, and Genetic Testing for BRCA- Related Cancer: Systematic Review to Update the U.S. Preventive Services Task Force Recommendation. Evidence Synthesis No. 101 (AHRQ Publication No. 12-05164-EF-1). Rockville, MD Agency for Healthcare Research and Quality; 2013.
- 24. Kuchenbaecker KB, Hopper JL, Barnes DR, et al. Risks of Breast, Ovarian, and Contralateral Breast Cancer for BRCA1 and BRCA2 Mutation Carriers. JAMA. Jun 20 2017; 317(23): 2402-2416. PMID 28632866

- 25. Begg CB. On the use of familial aggregation in population-based case probands for calculating penetrance. J Natl Cancer Inst. Aug 21 2002; 94(16): 1221-6. PMID 12189225
- 26. Thorlacius S, Struewing JP, Hartge P, et al. Population-based study of risk of breast cancer in carriers of BRCA2 mutation. Lancet. Oct 24 1998; 352(9137): 1337-9. PMID 9802270
- 27. King MC, Marks JH, Mandell JB. Breast and ovarian cancer risks due to inherited mutations in BRCA1 and BRCA2. Science. Oct 24 2003; 302(5645): 643-6. PMID 14576434
- 28. Metcalfe K, Lynch HT, Ghadirian P, et al. Contralateral breast cancer in BRCA1 and BRCA2 mutation carriers. J Clin Oncol. Jun 15 2004; 22(12): 2328-35. PMID 15197194
- 29. Mavaddat N, Peock S, Frost D, et al. Cancer risks for BRCA1 and BRCA2 mutation carriers: results from prospective analysis of EMBRACE. J Natl Cancer Inst. Jun 05 2013; 105(11): 812-22. PMID 23628597
- 30. Trainer AH, Meiser B, Watts K, et al. Moving toward personalized medicine: treatment-focused genetic testing of women newly diagnosed with ovarian cancer. Int J Gynecol Cancer. Jul 2010; 20(5): 704-16. PMID 20973257
- 31. Zhang S, Royer R, Li S, et al. Frequencies of BRCA1 and BRCA2 mutations among 1,342 unselected patients with invasive ovarian cancer. Gynecol Oncol. May 01 2011; 121(2): 353-7. PMID 21324516
- 32. Kurian AW, Hughes, E., Handorf, E. A., et al. Breast and ovarian cancer penetrance estimates derived from germline multiple-gene sequencing results in women. Precis Oncol. 2017;1:1-12.
- 33. Langer LR, McCoy H, Kidd J, et al. Hereditary cancer testing in patients with ovarian cancer using a 25-gene panel. J Community Supportive Oncol. 2016;14(7):314-319.
- 34. Norquist BM, Harrell MI, Brady MF, et al. Inherited Mutations in Women With Ovarian Carcinoma. JAMA Oncol. Apr 2016; 2(4): 482-90. PMID 26720728
- 35. Harter P, Hauke J, Heitz F, et al. Prevalence of deleterious germline variants in risk genes including BRCA1/2 in consecutive ovarian cancer patients (AGO-TR-1). PLoS One. 2017; 12(10): e0186043. PMID 29053726
- 36. Hirst JE, Gard GB, McIllroy K, et al. High rates of occult fallopian tube cancer diagnosed at prophylactic bilateral salpingo-oophorectomy. Int J Gynecol Cancer. Jul 2009; 19(5): 826-9. PMID 19574767
- 37. Powell CB, Swisher EM, Cass I, et al. Long term follow up of BRCA1 and BRCA2 mutation carriers with unsuspected neoplasia identified at risk reducing salpingo-oophorectomy. Gynecol Oncol. May 2013; 129(2): 364-71. PMID 23391663
- 38. Hruban RH, Canto MI, Goggins M, et al. Update on familial pancreatic cancer. Adv Surg. 2010; 44: 293-311. PMID 20919528
- 39. Couch FJ, Johnson MR, Rabe KG, et al. The prevalence of BRCA2 mutations in familial pancreatic cancer. Cancer Epidemiol Biomarkers Prev. Feb 2007; 16(2): 342-6. PMID 17301269
- 40. Ferrone CR, Levine DA, Tang LH, et al. BRCA germline mutations in Jewish patients with pancreatic adenocarcinoma. J Clin Oncol. Jan 20 2009; 27(3): 433-8. PMID 19064968
- 41. Holter S, Borgida A, Dodd A, et al. Germline BRCA Mutations in a Large Clinic-Based Cohort of Patients With Pancreatic Adenocarcinoma. J Clin Oncol. Oct 01 2015; 33(28): 3124-9. PMID 25940717

- 42. Shindo K, Yu J, Suenaga M, et al. Deleterious Germline Mutations in Patients With Apparently Sporadic Pancreatic Adenocarcinoma. J Clin Oncol. Oct 20 2017; 35(30): 3382-3390. PMID 28767289
- 43. Yurgelun MB, Chittenden AB, Morales-Oyarvide V, et al. Germline cancer susceptibility gene variants, somatic second hits, and survival outcomes in patients with resected pancreatic cancer. Genet Med. Jan 2019; 21(1): 213-223. PMID 29961768
- 44. Hu C, Hart SN, Polley EC, et al. Association Between Inherited Germline Mutations in Cancer Predisposition Genes and Risk of Pancreatic Cancer. JAMA. Jun 19 2018; 319(23): 2401-2409. PMID 29922827
- 45. Edwards SM, Kote-Jarai Z, Meitz J, et al. Two percent of men with early-onset prostate cancer harbor germline mutations in the BRCA2 gene. Am J Hum Genet. Jan 2003; 72(1): 1-12. PMID 12474142
- 46. Pritchard CC, Mateo J, Walsh MF, et al. Inherited DNA-Repair Gene Mutations in Men with Metastatic Prostate Cancer. N Engl J Med. Aug 04 2016; 375(5): 443-53. PMID 27433846
- 47. Abida W, Armenia J, Gopalan A, et al. Prospective Genomic Profiling of Prostate Cancer Across Disease States Reveals Germline and Somatic Alterations That May Affect Clinical Decision Making. JCO Precis Oncol. Jul 2017; 2017. PMID 28825054
- 48. Walsh T, Casadei S, Coats KH, et al. Spectrum of mutations in BRCA1, BRCA2, CHEK2, and TP53 in families at high risk of breast cancer. JAMA. Mar 22 2006; 295(12): 1379-88. PMID 16551709
- 49. Palma MD, Domchek SM, Stopfer J, et al. The relative contribution of point mutations and genomic rearrangements in BRCA1 and BRCA2 in high-risk breast cancer families. Cancer Res. Sep 01 2008; 68(17): 7006-14. PMID 18703817
- 50. Nelson HD, Pappas M, Cantor A, et al. Risk Assessment, Genetic Counseling, and Genetic Testing for BRCA-Related Cancer in Women: Updated Evidence Report and Systematic Review for the US Preventive Services Task Force. JAMA. Aug 20 2019; 322(7): 666-685. PMID 31429902
- 51. Grann VR, Whang W, Jacobson JS, et al. Benefits and costs of screening Ashkenazi Jewish women for BRCA1 and BRCA2. J Clin Oncol. Feb 1999; 17(2): 494-500. PMID 10080590
- 52. Hartmann LC, Schaid DJ, Woods JE, et al. Efficacy of bilateral prophylactic mastectomy in women with a family history of breast cancer. N Engl J Med. Jan 14 1999; 340(2): 77-84. PMID 9887158
- 53. Menkiszak J, Rzepka-Górska I, Górski B, et al. Attitudes toward preventive oophorectomy among BRCA1 mutation carriers in Poland. Eur J Gynaecol Oncol. 2004; 25(1): 93-5. PMID 15053071
- 54. Møller P, Borg A, Evans DG, et al. Survival in prospectively ascertained familial breast cancer: analysis of a series stratified by tumour characteristics, BRCA mutations and oophorectomy. Int J Cancer. Oct 20 2002; 101(6): 555-9. PMID 12237897
- 55. Olopade OI, Artioli G. Efficacy of risk-reducing salpingo-oophorectomy in women with BRCA-1 and BRCA-2 mutations. Breast J. 2004; 10 Suppl 1: S5-9. PMID 14984481
- 56. Rebbeck TR, Lynch HT, Neuhausen SL, et al. Prophylactic oophorectomy in carriers of BRCA1 or BRCA2 mutations. N Engl J Med. May 23 2002; 346(21): 1616-22. PMID 12023993

- 57. Weitzel JN, McCaffrey SM, Nedelcu R, et al. Effect of genetic cancer risk assessment on surgical decisions at breast cancer diagnosis. Arch Surg. Dec 2003; 138(12): 1323-8; discussion 1329. PMID 14662532
- 58. Finch AP, Lubinski J, Møller P, et al. Impact of oophorectomy on cancer incidence and mortality in women with a BRCA1 or BRCA2 mutation. J Clin Oncol. May 20 2014; 32(15): 1547-53. PMID 24567435
- 59. Domchek SM, Friebel TM, Singer CF, et al. Association of risk-reducing surgery in BRCA1 or BRCA2 mutation carriers with cancer risk and mortality. JAMA. Sep 01 2010; 304(9): 967-75. PMID 20810374
- 60. Elmi M, Azin A, Elnahas A, et al. Concurrent risk-reduction surgery in patients with increased lifetime risk for breast and ovarian cancer: an analysis of the National Surgical Quality Improvement Program (NSQIP) database. Breast Cancer Res Treat. Aug 2018; 171(1): 217-223. PMID 29761322
- 61. Li X, You R, Wang X, et al. Effectiveness of Prophylactic Surgeries in BRCA1 or BRCA2 Mutation Carriers: A Meta-analysis and Systematic Review. Clin Cancer Res. Aug 01 2016; 22(15): 3971-81. PMID 26979395
- 62. Ludwig KK, Neuner J, Butler A, et al. Risk reduction and survival benefit of prophylactic surgery in BRCA mutation carriers, a systematic review. Am J Surg. Oct 2016; 212(4): 660-669. PMID 27649974
- 63. Marchetti C, De Felice F, Palaia I, et al. Risk-reducing salpingo-oophorectomy: a metaanalysis on impact on ovarian cancer risk and all cause mortality in BRCA 1 and BRCA 2 mutation carriers. BMC Womens Health. Dec 12 2014; 14: 150. PMID 25494812
- 64. Scheuer L, Kauff N, Robson M, et al. Outcome of preventive surgery and screening for breast and ovarian cancer in BRCA mutation carriers. J Clin Oncol. Mar 01 2002; 20(5): 1260-8. PMID 11870168
- 65. Mitra AV, Bancroft EK, Barbachano Y, et al. Targeted prostate cancer screening in men with mutations in BRCA1 and BRCA2 detects aggressive prostate cancer: preliminary analysis of the results of the IMPACT study. BJU Int. Jan 2011; 107(1): 28-39. PMID 20840664
- 66. Suszynska M, Klonowska K, Jasinska AJ, et al. Large-scale meta-analysis of mutations identified in panels of breast/ovarian cancer-related genes Providing evidence of cancer predisposition genes. Gynecol Oncol. May 2019; 153(2): 452-462. PMID 30733081
- 67. Erkko H, Dowty JG, Nikkilä J, et al. Penetrance analysis of the PALB2 c.1592delT founder mutation. Clin Cancer Res. Jul 15 2008; 14(14): 4667-71. PMID 18628482
- 68. Heikkinen T, Kärkkäinen H, Aaltonen K, et al. The breast cancer susceptibility mutation PALB2 1592delT is associated with an aggressive tumor phenotype. Clin Cancer Res. May 01 2009; 15(9): 3214-22. PMID 19383810
- 69. Rahman N, Seal S, Thompson D, et al. PALB2, which encodes a BRCA2-interacting protein, is a breast cancer susceptibility gene. Nat Genet. Feb 2007; 39(2): 165-7. PMID 17200668
- 70. Thompson ER, Gorringe KL, Rowley SM, et al. Prevalence of PALB2 mutations in Australian familial breast cancer cases and controls. Breast Cancer Res. Aug 19 2015; 17(1): 111. PMID 26283626
- 71. Southey MC, Goldgar DE, Winqvist R, et al. PALB2, CHEK2 and ATM rare variants and cancer risk: data from COGS. J Med Genet. Dec 2016; 53(12): 800-811. PMID 27595995

- 72. Lu HM, Li S, Black MH, et al. Association of Breast and Ovarian Cancers With Predisposition Genes Identified by Large-Scale Sequencing. JAMA Oncol. Jan 01 2019; 5(1): 51-57. PMID 30128536
- 73. Woodward ER, van Veen EM, Forde C, et al. Clinical utility of testing for PALB2 and CHEK2 c.1100delC in breast and ovarian cancer. Genet Med. Oct 2021; 23(10): 1969-1976. PMID 34113003
- 74. Yang X, Leslie G, Doroszuk A, et al. Cancer Risks Associated With Germline PALB2 Pathogenic Variants: An International Study of 524 Families. J Clin Oncol. Mar 01 2020; 38(7): 674-685. PMID 31841383
- 75. Li N, Lim BWX, Thompson ER, et al. Investigation of monogenic causes of familial breast cancer: data from the BEACCON case-control study. NPJ Breast Cancer. Jun 11 2021; 7(1): 76. PMID 34117267
- 76. Antoniou AC, Foulkes WD, Tischkowitz M. Breast cancer risk in women with PALB2 mutations in different populations. Lancet Oncol. Aug 2015; 16(8): e375-6. PMID 26248842
- 77. National Cancer Institute, Surveillance Epidemiology and End Results Program. Cancer Stat Facts: Female Breast Cancer. n.d.; https://seer.cancer.gov/statfacts/html/breast.html. Accessed June 26, 2024.
- 78. Rosenthal ET, Evans B, Kidd J, et al. Increased Identification of Candidates for High-Risk Breast Cancer Screening Through Expanded Genetic Testing. J Am Coll Radiol. Apr 2017; 14(4): 561-568. PMID 28011157
- 79. Phi XA, Saadatmand S, De Bock GH, et al. Contribution of mammography to MRI screening in BRCA mutation carriers by BRCA status and age: individual patient data meta-analysis. Br J Cancer. Mar 15 2016; 114(6): 631-7. PMID 26908327
- 80. Phillips KA, Milne RL, Rookus MA, et al. Tamoxifen and risk of contralateral breast cancer for BRCA1 and BRCA2 mutation carriers. J Clin Oncol. Sep 01 2013; 31(25): 3091-9. PMID 23918944
- 81. Hartmann LC, Sellers TA, Schaid DJ, et al. Efficacy of bilateral prophylactic mastectomy in BRCA1 and BRCA2 gene mutation carriers. J Natl Cancer Inst. Nov 07 2001; 93(21): 1633-7. PMID 11698567
- 82. Portschy PR, Kuntz KM, Tuttle TM. Survival outcomes after contralateral prophylactic mastectomy: a decision analysis. J Natl Cancer Inst. Aug 2014; 106(8). PMID 25031308
- 83. Schrag D, Kuntz KM, Garber JE, et al. Decision analysis--effects of prophylactic mastectomy and oophorectomy on life expectancy among women with BRCA1 or BRCA2 mutations. N Engl J Med. May 15 1997; 336(20): 1465-71. PMID 9148160
- 84. Schrag D, Kuntz KM, Garber JE, et al. Life expectancy gains from cancer prevention strategies for women with breast cancer and BRCA1 or BRCA2 mutations. JAMA. Feb 02 2000; 283(5): 617-24. PMID 10665701
- 85. Bedrosian I, Somerfield MR, Achatz MI, et al. Germline Testing in Patients With Breast Cancer: ASCO-Society of Surgical Oncology Guideline. J Clin Oncol. Feb 10 2024; 42(5): 584-604. PMID 38175972
- 86. Tung N, Ricker C, Messersmith H, et al. Selection of Germline Genetic Testing Panels in Patients With Cancer: ASCO Guideline. J Clin Oncol. Jul 20 2024; 42(21): 2599-2615. PMID 38759122
- 87. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Genetic/Familial High Risk Assessment: Breast, Ovarian, and Pancreatic. Version 3.2024.

- https://www.nccn.org/professionals/physician_gls/pdf/genetics_bop.pdf. Accessed June 26, 2024.
- 88. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Ovarian Cancer. Version 2.2024. https://www.nccn.org/professionals/physician_gls/pdf/ovarian.pdf. Accessed June 25, 2024.
- 89. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Pancreatic Adenocarcinoma. Version 2.2024. https://www.nccn.org/professionals/physician_gls/pdf/pancreatic.pdf. Accessed June 24, 2024.
- 90. National Comprehensive Cancer Network Clinical Practice Guidelines in Oncology: Neuroendocrine and Adrenal Tumors Version 2.2022
- 91. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Prostate Cancer. Version 4.2024. https://www.nccn.org/professionals/physician_gls/pdf/prostate.pdf. Accessed June 23, 2024.
- 92. The American Society of Breast Surgeons. Consensus Guidelines on Genetic Testing for Hereditary Breast Cancer. 2019. https://www.breastsurgeons.org/docs/statements/Consensus-Guideline-on-Genetic-Testing-for-Hereditary-Breast-Cancer.pdf. Accessed June 26, 2024
- 93. Lancaster JM, Powell CB, Chen LM, et al. Society of Gynecologic Oncology statement on risk assessment for inherited gynecologic cancer predispositions. Gynecol Oncol. Jan 2015; 136(1): 3-7. PMID 25238946
- 94. Practice Bulletin No. 182 Summary: Hereditary Breast and Ovarian Cancer Syndrome. Obstet Gynecol. Sep 2017; 130(3): 657-659. PMID 28832475
- 95. Owens DK, Davidson KW, Krist AH, et al. Risk Assessment, Genetic Counseling, and Genetic Testing for BRCA-Related Cancer: US Preventive Services Task Force Recommendation Statement. JAMA. Aug 20 2019; 322(7): 652-665. PMID 31429903

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- 1. Vogt S, Jones N, Christian D, et al. Expanded extracolonic tumor spectrum in MUTYH-associated polyposis. Gastroenterology. Dec 2009; 137(6): 1976-85.e1-10. PMID 19732775
- 2. Balmaña J, Castells A, Cervantes A. Familial colorectal cancer risk: ESMO Clinical Practice Guidelines. Ann Oncol. May 2010; 21 Suppl 5: v78-81. PMID 20555108
- 3. Gala M, Chung DC. Hereditary colon cancer syndromes. Semin Oncol. Aug 2011; 38(4): 490-9. PMID 21810508
- 4. Quehenberger F, Vasen HF, van Houwelingen HC. Risk of colorectal and endometrial cancer for carriers of mutations of the hMLH1 and hMSH2 gene: correction for ascertainment. J Med Genet. Jun 2005; 42(6): 491-6. PMID 15937084
- 5. Guindalini RS, Win AK, Gulden C, et al. Mutation spectrum and risk of colorectal cancer in African American families with Lynch syndrome. Gastroenterology. Nov 2015; 149(6): 1446-53. PMID 26248088
- 6. Sinn DH, Chang DK, Kim YH, et al. Effectiveness of each Bethesda marker in defining microsatellite instability when screening for Lynch syndrome. Hepatogastroenterology. 2009; 56(91-92): 672-6. PMID 19621678

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- 7. Wu Y, Berends MJ, Mensink RG, et al. Association of hereditary nonpolyposis colorectal cancer-related tumors displaying low microsatellite instability with MSH6 germline mutations. Am J Hum Genet. Nov 1999; 65(5): 1291-8. PMID 10521294
- 8. Goel A, Nagasaka T, Spiegel J, et al. Low frequency of Lynch syndrome among young patients with non-familial colorectal cancer. Clin Gastroenterol Hepatol. Nov 2010; 8(11): 966-71. PMID 20655395
- 9. Palomaki GE, McClain MR, Melillo S, et al. EGAPP supplementary evidence review: DNA testing strategies aimed at reducing morbidity and mortality from Lynch syndrome. Genet Med. Jan 2009; 11(1): 42-65. PMID 19125127
- 10. Bouzourene H, Hutter P, Losi L, et al. Selection of patients with germline MLH1 mutated Lynch syndrome by determination of MLH1 methylation and BRAF mutation. Fam Cancer. Jun 2010; 9(2): 167-72. PMID 19949877
- 11. Niessen RC, Hofstra RM, Westers H, et al. Germline hypermethylation of MLH1 and EPCAM deletions are a frequent cause of Lynch syndrome. Genes Chromosomes Cancer. Aug 2009; 48(8): 737-44. PMID 19455606
- 12. Hesson LB, Hitchins MP, Ward RL. Epimutations and cancer predisposition: importance and mechanisms. Curr Opin Genet Dev. Jun 2010; 20(3): 290-8. PMID 20359882
- 13. Hitchins MP. Inheritance of epigenetic aberrations (constitutional epimutations) in cancer susceptibility. Adv Genet. 2010; 70: 201-43. PMID 20920750
- 14. Vasen HF, Watson P, Mecklin JP, et al. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. Gastroenterology. Jun 1999; 116(6): 1453-6. PMID 10348829
- 15. Umar A, Boland CR, Terdiman JP, et al. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. J Natl Cancer Inst. Feb 18 2004; 96(4): 261-8. PMID 14970275
- 16. Kastrinos F, Uno H, Ukaegbu C, et al. Development and Validation of the PREMM 5 Model for Comprehensive Risk Assessment of Lynch Syndrome. J Clin Oncol. Jul 01 2017; 35(19): 2165-2172. PMID 28489507
- 17. Latchford AR, Neale K, Phillips RK, et al. Juvenile polyposis syndrome: a study of genotype, phenotype, and long-term outcome. Dis Colon Rectum. Oct 2012; 55(10): 1038-43. PMID 22965402
- 18. Howe JR, Roth S, Ringold JC, et al. Mutations in the SMAD4/DPC4 gene in juvenile polyposis. Science. May 15 1998; 280(5366): 1086-8. PMID 9582123
- 19. Fogt F, Brown CA, Badizadegan K, et al. Low prevalence of loss of heterozygosity and SMAD4 mutations in sporadic and familial juvenile polyposis syndrome-associated juvenile polyps. Am J Gastroenterol. Oct 2004; 99(10): 2025-31. PMID 15447767
- 20. Burger B, Uhlhaas S, Mangold E, et al. Novel de novo mutation of MADH4/SMAD4 in a patient with juvenile polyposis. Am J Med Genet. Jul 01 2002; 110(3): 289-91. PMID 12116240
- 21. Syngal S, Brand RE, Church JM, et al. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. Am J Gastroenterol. Feb 2015; 110(2): 223-62; quiz 263. PMID 25645574
- 22. Grotsky HW, Rickert RR, Smith WD, et al. Familial juvenile polyposis coli. A clinical and pathologic study of a large kindred. Gastroenterology. Mar 1982; 82(3): 494-501. PMID 7054044

- 23. Schreibman IR, Baker M, Amos C, et al. The hamartomatous polyposis syndromes: a clinical and molecular review. Am J Gastroenterol. Feb 2005; 100(2): 476-90. PMID 15667510
- 24. Brosens LA, van Hattem A, Hylind LM, et al. Risk of colorectal cancer in juvenile polyposis. Gut. Jul 2007; 56(7): 965-7. PMID 17303595
- 25. Gallione CJ, Repetto GM, Legius E, et al. A combined syndrome of juvenile polyposis and hereditary haemorrhagic telangiectasia associated with mutations in MADH4 (SMAD4). Lancet. Mar 13 2004; 363(9412): 852-9. PMID 15031030
- 26. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Colon Cancer. Version 4.2024. http://www.nccn.org/professionals/physician_gls/pdf/colon.pdf. Accessed July 10, 2024
- 27. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Uterine Neoplasms. Version 2.2024. https://www.nccn.org/professionals/physician_gls/pdf/uterine.pdf Accessed July 13, 2024.
- 28. Olschwang S, Markie D, Seal S, et al. Peutz-Jeghers disease: most, but not all, families are compatible with linkage to 19p13.3. J Med Genet. Jan 1998; 35(1): 42-4. PMID 9475093
- 29. Jenne DE, Reimann H, Nezu J, et al. Peutz-Jeghers syndrome is caused by mutations in a novel serine threonine kinase. Nat Genet. Jan 1998; 18(1): 38-43. PMID 9425897
- 30. Hemminki A, Markie D, Tomlinson I, et al. A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. Nature. Jan 08 1998; 391(6663): 184-7. PMID 9428765
- 31. Hernan I, Roig I, Martin B, et al. De novo germline mutation in the serine-threonine kinase STK11/LKB1 gene associated with Peutz-Jeghers syndrome. Clin Genet. Jul 2004; 66(1): 58-62. PMID 15200509
- 32. van Lier MG, Wagner A, Mathus-Vliegen EM, et al. High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations. Am J Gastroenterol. Jun 2010; 105(6): 1258-64; author reply 1265. PMID 20051941
- 33. Blue Cross and Blue Shield Association Technology Evaluation Center (TEC). Genetic Testing for Inherited Susceptibility to Colorectal Cancer: Part I Adenomatous Polyposis Coli Gene Mutations. TEC Assessments. 1998; Volume 13:Tab 10. PMID
- 34. Kastrinos F, Syngal S. Recently identified colon cancer predispositions: MYH and MSH6 mutations. Semin Oncol. Oct 2007; 34(5): 418-24. PMID 17920897
- 35. Lefevre JH, Parc Y, Svrcek M, et al. APC, MYH, and the correlation genotype-phenotype in colorectal polyposis. Ann Surg Oncol. Apr 2009; 16(4): 871-7. PMID 19169759
- 36. Avezzù A, Agostini M, Pucciarelli S, et al. The role of MYH gene in genetic predisposition to colorectal cancer: another piece of the puzzle. Cancer Lett. Sep 18 2008; 268(2): 308-13. PMID 18495334
- 37. Balaguer F, Castellví-Bel S, Castells A, et al. Identification of MYH mutation carriers in colorectal cancer: a multicenter, case-control, population-based study. Clin Gastroenterol Hepatol. Mar 2007; 5(3): 379-87. PMID 17368238
- 38. Jasperson KW, Patel SG, Ahnen DJ. APC-Associated Polyposis Conditions. In: Pagon RA, Adam MP, Ardinger HH, et al., eds. GeneReviews. Seattle, WA: University of Washington; 2017.

- 39. Lagarde A, Rouleau E, Ferrari A, et al. Germline APC mutation spectrum derived from 863 genomic variations identified through a 15-year medical genetics service to French patients with FAP. J Med Genet. Oct 2010; 47(10): 721-2. PMID 20685668
- 40. Aretz S, Stienen D, Uhlhaas S, et al. Large submicroscopic genomic APC deletions are a common cause of typical familial adenomatous polyposis. J Med Genet. Feb 2005; 42(2): 185-92. PMID 15689459
- 41. Bunyan DJ, Eccles DM, Sillibourne J, et al. Dosage analysis of cancer predisposition genes by multiplex ligation-dependent probe amplification. Br J Cancer. Sep 13 2004; 91(6): 1155-9. PMID 15475941
- 42. Out AA, Tops CM, Nielsen M, et al. Leiden Open Variation Database of the MUTYH gene. Hum Mutat. Nov 2010; 31(11): 1205-15. PMID 20725929
- 43. Nielsen M, Lynch H, Infante E, et al. MUTYH-Associated Polyposis. In: Pagon RA, Adam MP, Ardinger HH, eds. GeneReviews Seattle, WA: University of Washington; 2012.
- 44. Sieber OM, Lamlum H, Crabtree MD, et al. Whole-gene APC deletions cause classical familial adenomatous polyposis, but not attenuated polyposis or "multiple" colorectal adenomas. Proc Natl Acad Sci U S A. Mar 05 2002; 99(5): 2954-8. PMID 11867715
- 45. Aretz S, Uhlhaas S, Goergens H, et al. MUTYH-associated polyposis: 70 of 71 patients with biallelic mutations present with an attenuated or atypical phenotype. Int J Cancer. Aug 15 2006; 119(4): 807-14. PMID 16557584
- 46. Michils G, Tejpar S, Thoelen R, et al. Large deletions of the APC gene in 15% of mutation-negative patients with classical polyposis (FAP): a Belgian study. Hum Mutat. Feb 2005; 25(2): 125-34. PMID 15643602
- 47. Truta B, Allen BA, Conrad PG, et al. A comparison of the phenotype and genotype in adenomatous polyposis patients with and without a family history. Fam Cancer. 2005; 4(2): 127-33. PMID 15951963
- 48. Bonis PA, Trikalinos TA, Chung M, et al. Hereditary Nonpolyposis Colorectal Cancer: Diagnostic Strategies and Their Implications (Evidence Report/Technology Assessment No. 150). Rockville, MD: Agency for Healthcare Research and Quality; 2007.
- 49. Berg AO, Armstrong K, Botkin J, et al. Recommendations from the EGAPP Working Group: genetic testing strategies in newly diagnosed individuals with colorectal cancer aimed at reducing morbidity and mortality from Lynch syndrome in relatives. Genet Med. Jan 2009; 11(1): 35-41. PMID 19125126
- 50. Vos JR, Fakkert IE, Spruijt L, et al. Evaluation of yield and experiences of age-related molecular investigation for heritable and nonheritable causes of mismatch repair deficient colorectal cancer to identify Lynch syndrome. Int J Cancer. Oct 15 2020; 147(8): 2150-2158. PMID 32510614
- 51. Tsuruta T, Todo Y, Yamada R, et al. Initial screening by immunohistochemistry is effective in universal screening for Lynch syndrome in endometrial cancer patients: a prospective observational study. Jpn J Clin Oncol. Jul 08 2022; 52(7): 752-758. PMID 35438162
- 52. Kloor M, Voigt AY, Schackert HK, et al. Analysis of EPCAM protein expression in diagnostics of Lynch syndrome. J Clin Oncol. Jan 10 2011; 29(2): 223-7. PMID 21115857
- 53. Kuiper RP, Vissers LE, Venkatachalam R, et al. Recurrence and variability of germline EPCAM deletions in Lynch syndrome. Hum Mutat. Apr 2011; 32(4): 407-14. PMID 21309036

- 54. Kovacs ME, Papp J, Szentirmay Z, et al. Deletions removing the last exon of TACSTD1 constitute a distinct class of mutations predisposing to Lynch syndrome. Hum Mutat. Feb 2009; 30(2): 197-203. PMID 19177550
- 55. Ligtenberg MJ, Kuiper RP, Chan TL, et al. Heritable somatic methylation and inactivation of MSH2 in families with Lynch syndrome due to deletion of the 3' exons of TACSTD1. Nat Genet. Jan 2009; 41(1): 112-7. PMID 19098912
- 56. Rumilla K, Schowalter KV, Lindor NM, et al. Frequency of deletions of EPCAM (TACSTD1) in MSH2-associated Lynch syndrome cases. J Mol Diagn. Jan 2011; 13(1): 93-9. PMID 21227399
- 57. Kempers MJ, Kuiper RP, Ockeloen CW, et al. Risk of colorectal and endometrial cancers in EPCAM deletion-positive Lynch syndrome: a cohort study. Lancet Oncol. Jan 2011; 12(1): 49-55. PMID 21145788
- 58. Jin M, Hampel H, Zhou X, et al. BRAF V600E mutation analysis simplifies the testing algorithm for Lynch syndrome. Am J Clin Pathol. Aug 2013; 140(2): 177-83. PMID 23897252
- 59. Capper D, Voigt A, Bozukova G, et al. BRAF V600E-specific immunohistochemistry for the exclusion of Lynch syndrome in MSI-H colorectal cancer. Int J Cancer. Oct 01 2013; 133(7): 1624-30. PMID 23553055
- 60. Kastrinos F, Syngal S. Screening patients with colorectal cancer for Lynch syndrome: what are we waiting for?. J Clin Oncol. Apr 01 2012; 30(10): 1024-7. PMID 22355054
- 61. Hampel H, Frankel WL, Martin E, et al. Screening for the Lynch syndrome (hereditary nonpolyposis colorectal cancer). N Engl J Med. May 05 2005; 352(18): 1851-60. PMID 15872200
- 62. Aktan-Collan K, Mecklin JP, Järvinen H, et al. Predictive genetic testing for hereditary non-polyposis colorectal cancer: uptake and long-term satisfaction. Int J Cancer. Jan 20 2000; 89(1): 44-50. PMID 10719730
- 63. Aktan-Collan K, Haukkala A, Pylvänäinen K, et al. Direct contact in inviting high-risk members of hereditary colon cancer families to genetic counselling and DNA testing. J Med Genet. Nov 2007; 44(11): 732-8. PMID 17630403
- 64. Stanley AJ, Gaff CL, Aittomäki AK, et al. Value of predictive genetic testing in management of hereditary non-polyposis colorectal cancer (HNPCC). Med J Aust. Apr 03 2000; 172(7): 313-6. PMID 10844916
- 65. Hadley DW, Jenkins J, Dimond E, et al. Genetic counseling and testing in families with hereditary nonpolyposis colorectal cancer. Arch Intern Med. Mar 10 2003; 163(5): 573-82. PMID 12622604
- 66. Lerman C, Hughes C, Trock BJ, et al. Genetic testing in families with hereditary nonpolyposis colon cancer. JAMA. May 05 1999; 281(17): 1618-22. PMID 10235155
- 67. Codori AM, Petersen GM, Miglioretti DL, et al. Attitudes toward colon cancer gene testing: factors predicting test uptake. Cancer Epidemiol Biomarkers Prev. Apr 1999; 8(4 Pt 2): 345-51. PMID 10207639
- 68. Schmeler KM, Lynch HT, Chen LM, et al. Prophylactic surgery to reduce the risk of gynecologic cancers in the Lynch syndrome. N Engl J Med. Jan 19 2006; 354(3): 261-9. PMID 16421367
- 69. Burke W, Petersen G, Lynch P, et al. Recommendations for follow-up care of individuals with an inherited predisposition to cancer. I. Hereditary nonpolyposis colon cancer. Cancer Genetics Studies Consortium. JAMA. Mar 19 1997; 277(11): 915-9. PMID 9062331

- 70. Guillem JG, Wood WC, Moley JF, et al. ASCO/SSO review of current role of risk-reducing surgery in common hereditary cancer syndromes. J Clin Oncol. Oct 01 2006; 24(28): 4642-60. PMID 17008706
- 71. Bonadona V, Bonaïti B, Olschwang S, et al. Cancer risks associated with germline mutations in MLH1, MSH2, and MSH6 genes in Lynch syndrome. JAMA. Jun 08 2011; 305(22): 2304-10. PMID 21642682
- 72. Obermair A, Youlden DR, Young JP, et al. Risk of endometrial cancer for women diagnosed with HNPCC-related colorectal carcinoma. Int J Cancer. Dec 01 2010; 127(11): 2678-84. PMID 20533284
- 73. Auranen A, Joutsiniemi T. A systematic review of gynecological cancer surveillance in women belonging to hereditary nonpolyposis colorectal cancer (Lynch syndrome) families. Acta Obstet Gynecol Scand. May 2011; 90(5): 437-44. PMID 21306348
- 74. Calva-Cerqueira D, Chinnathambi S, Pechman B, et al. The rate of germline mutations and large deletions of SMAD4 and BMPR1A in juvenile polyposis. Clin Genet. Jan 2009; 75(1): 79-85. PMID 18823382
- 75. Aretz S, Stienen D, Uhlhaas S, et al. High proportion of large genomic deletions and a genotype phenotype update in 80 unrelated families with juvenile polyposis syndrome. J Med Genet. Nov 2007; 44(11): 702-9. PMID 17873119
- 76. Volikos E, Robinson J, Aittomäki K, et al. LKB1 exonic and whole gene deletions are a common cause of Peutz-Jeghers syndrome. J Med Genet. May 2006; 43(5): e18. PMID 16648371
- 77. Aretz S, Stienen D, Uhlhaas S, et al. High proportion of large genomic STK11 deletions in Peutz-Jeghers syndrome. Hum Mutat. Dec 2005; 26(6): 513-9. PMID 16287113
- 78. Kastrinos F, Steyerberg EW, Mercado R, et al. The PREMM(1,2,6) model predicts risk of MLH1, MSH2, and MSH6 germline mutations based on cancer history. Gastroenterology. Jan 2011; 140(1): 73-81. PMID 20727894
- 79. Stoffel EM, Mangu PB, Gruber SB, et al. Hereditary colorectal cancer syndromes: American Society of Clinical Oncology Clinical Practice Guideline endorsement of the familial risk-colorectal cancer: European Society for Medical Oncology Clinical Practice Guidelines. J Clin Oncol. Jan 10 2015; 33(2): 209-17. PMID 25452455
- 80. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Genetic/Familial High-Risk Assessment: Colorectal. Version 2.2023. http://www.nccn.org/professionals/physician_gls/pdf/genetics_colon.pdf. Accessed July 12, 2024.
- 81. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Colorectal Cancer Screening. Version 1. 2024. http://www.nccn.org/professionals/physician_gls/pdf/colorectal_screening.pdf. Accessed July 11, 2024.

JAK2, MPL, and CALR Testing for Myeloproliferative Neoplasms

- 1. Jones AV, Kreil S, Zoi K, et al. Widespread occurrence of the JAK2 V617F mutation in chronic myeloproliferative disorders. Blood. Sep 15 2005; 106(6): 2162-8. PMID 15920007
- 2. Murphy S, Peterson P, Iland H, et al. Experience of the Polycythemia Vera Study Group with essential thrombocythemia: a final report on diagnostic criteria, survival, and leukemic transition by treatment. Semin Hematol. Jan 1997; 34(1): 29-39. PMID 9025160

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- 3. Pearson TC, Messinezy M. The diagnostic criteria of polycythaemia rubra vera. Leuk Lymphoma. Sep 1996; 22 Suppl 1: 87-93. PMID 8951778
- 4. Vardiman JW, Harris NL, Brunning RD. The World Health Organization (WHO) classification of the myeloid neoplasms. Blood. Oct 01 2002; 100(7): 2292-302. PMID 12239137
- 5. Vardiman JW, Thiele J, Arber DA, et al. The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: rationale and important changes. Blood. Jul 30 2009; 114(5): 937-51. PMID 19357394
- 6. Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. Blood. May 19 2016; 127(20): 2391-405. PMID 27069254
- 7. Khoury JD, Solary E, Abla O, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Myeloid and Histiocytic/Dendritic Neoplasms. Leukemia. Jul 2022; 36(7): 1703-1719. PMID 35732831
- 8. Arber DA, Orazi A, Hasserjian RP, et al. International Consensus Classification of Myeloid Neoplasms and Acute Leukemias: integrating morphologic, clinical, and genomic data. Blood. Sep 15 2022; 140(11): 1200-1228. PMID 35767897
- 9. Tefferi A, Strand JJ, Lasho TL, et al. Bone marrow JAK2V617F allele burden and clinical correlates in polycythemia vera. Leukemia. Sep 2007; 21(9): 2074-5. PMID 17476276
- 10. Wilkins BS, Erber WN, Bareford D, et al. Bone marrow pathology in essential thrombocythemia: interobserver reliability and utility for identifying disease subtypes. Blood. Jan 01 2008; 111(1): 60-70. PMID 17885079
- 11. Baxter EJ, Scott LM, Campbell PJ, et al. Acquired mutation of the tyrosine kinase JAK2 in human myeloproliferative disorders. Lancet. Mar 2005; 365(9464): 1054-61. PMID 15781101
- 12. NIH Genetics Home Reference. JAK2 gene: Janus kinase 2. 2014. https://ghr.nlm.nih.gov/gene/JAK2. Accessed June 25, 2024.
- 13. Levine RL, Wadleigh M, Cools J, et al. Activating mutation in the tyrosine kinase JAK2 in polycythemia vera, essential thrombocythemia, and myeloid metaplasia with myelofibrosis. Cancer Cell. Apr 2005; 7(4): 387-97. PMID 15837627
- 14. James C, Ugo V, Le Couédic JP, et al. A unique clonal JAK2 mutation leading to constitutive signalling causes polycythaemia vera. Nature. Apr 28 2005; 434(7037): 1144-8. PMID 15793561
- 15. Kralovics R, Passamonti F, Buser AS, et al. A gain-of-function mutation of JAK2 in myeloproliferative disorders. N Engl J Med. Apr 28 2005; 352(17): 1779-90. PMID 15858187
- 16. Tefferi A, Sirhan S, Lasho TL, et al. Concomitant neutrophil JAK2 mutation screening and PRV-1 expression analysis in myeloproliferative disorders and secondary polycythaemia. Br J Haematol. Oct 2005; 131(2): 166-71. PMID 16197445
- 17. Zhao R, Xing S, Li Z, et al. Identification of an acquired JAK2 mutation in polycythemia vera. J Biol Chem. Jun 17 2005; 280(24): 22788-92. PMID 15863514
- 18. Campbell PJ, Scott LM, Buck G, et al. Definition of subtypes of essential thrombocythaemia and relation to polycythaemia vera based on JAK2 V617F mutation status: a prospective study. Lancet. Dec 03 2005; 366(9501): 1945-53. PMID 16325696

- 19. Wolanskyj AP, Lasho TL, Schwager SM, et al. JAK2 mutation in essential thrombocythaemia: clinical associations and long-term prognostic relevance. Br J Haematol. Oct 2005; 131(2): 208-13. PMID 16197451
- 20. Campbell PJ, Griesshammer M, Döhner K, et al. V617F mutation in JAK2 is associated with poorer survival in idiopathic myelofibrosis. Blood. Mar 01 2006; 107(5): 2098-100. PMID 16293597
- 21. Tefferi A, Lasho TL, Schwager SM, et al. The JAK2(V617F) tyrosine kinase mutation in myelofibrosis with myeloid metaplasia: lineage specificity and clinical correlates. Br J Haematol. Nov 2005; 131(3): 320-8. PMID 16225651
- 22. Xu X, Zhang Q, Luo J, et al. JAK2(V617F): Prevalence in a large Chinese hospital population. Blood. Jan 01 2007; 109(1): 339-42. PMID 16946305
- 23. Sidon P, El Housni H, Dessars B, et al. The JAK2V617F mutation is detectable at very low level in peripheral blood of healthy donors. Leukemia. Sep 2006; 20(9): 1622. PMID 16775613
- 24. Scott LM, Tong W, Levine RL, et al. JAK2 exon 12 mutations in polycythemia vera and idiopathic erythrocytosis. N Engl J Med. Feb 01 2007; 356(5): 459-68. PMID 17267906
- 25. Pardanani A, Lasho TL, Finke C, et al. Prevalence and clinicopathologic correlates of JAK2 exon 12 mutations in JAK2V617F-negative polycythemia vera. Leukemia. Sep 2007; 21(9): 1960-3. PMID 17597810
- 26. Siemiatkowska A, Bieniaszewska M, Hellmann A, et al. JAK2 and MPL gene mutations in V617F-negative myeloproliferative neoplasms. Leuk Res. Mar 2010; 34(3): 387-9. PMID 19643476
- 27. Cazzola M, Kralovics R. From Janus kinase 2 to calreticulin: the clinically relevant genomic landscape of myeloproliferative neoplasms. Blood. Jun 12 2014; 123(24): 3714-9. PMID 24786775
- 28. Makarik TV, Abdullaev AO, Nikulina EE, et al. Low JAK2 V617F Allele Burden in Ph-Negative Chronic Myeloproliferative Neoplasms Is Associated with Additional CALR or MPL Gene Mutations. Genes (Basel). Apr 12 2021; 12(4). PMID 33921387
- 29. Mejía-Ochoa M, Acevedo Toro PA, Cardona-Arias JA. Systematization of analytical studies of polycythemia vera, essential thrombocythemia and primary myelofibrosis, and a meta-analysis of the frequency of JAK2, CALR and MPL mutations: 2000-2018. BMC Cancer. Jun 17 2019; 19(1): 590. PMID 31208359
- 30. Kumar C, Purandare AV, Lee FY, et al. Kinase drug discovery approaches in chronic myeloproliferative disorders. Oncogene. Jun 18 2009; 28(24): 2305-13. PMID 19421140
- 31. Verstovsek S, Kantarjian H, Mesa RA, et al. Safety and efficacy of INCB018424, a JAK1 and JAK2 inhibitor, in myelofibrosis. N Engl J Med. Sep 16 2010; 363(12): 1117-27. PMID 20843246
- 32. Rambaldi A, Dellacasa CM, Finazzi G, et al. A pilot study of the Histone-Deacetylase inhibitor Givinostat in patients with JAK2V617F positive chronic myeloproliferative neoplasms. Br J Haematol. Aug 2010; 150(4): 446-55. PMID 20560970
- 33. Santos FP, Kantarjian HM, Jain N, et al. Phase 2 study of CEP-701, an orally available JAK2 inhibitor, in patients with primary or post-polycythemia vera/essential thrombocythemia myelofibrosis. Blood. Feb 11 2010; 115(6): 1131-6. PMID 20008298
- 34. Quintás-Cardama A, Verstovsek S. Spleen deflation and beyond: the pros and cons of Janus kinase 2 inhibitor therapy for patients with myeloproliferative neoplasms. Cancer. Feb 15 2012; 118(4): 870-7. PMID 21766300

- 35. Verstovsek S, Mesa RA, Gotlib J, et al. A double-blind, placebo-controlled trial of ruxolitinib for myelofibrosis. N Engl J Med. Mar 01 2012; 366(9): 799-807. PMID 22375971
- 36. Harrison C, Kiladjian JJ, Al-Ali HK, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. N Engl J Med. Mar 01 2012; 366(9): 787-98. PMID 22375970
- 37. Verstovsek S, Mesa RA, Gotlib J, et al. Efficacy, safety, and survival with ruxolitinib in patients with myelofibrosis: results of a median 3-year follow-up of COMFORT-I. Haematologica. Apr 2015; 100(4): 479-88. PMID 25616577
- 38. National Comprehensive Cancer Network (NCCN). NCCN Clinical Practice Guidelines in Oncology: Myeloproliferative neoplasms. Version.1.2024. https://www.nccn.org/professionals/physician_gls/pdf/mpn.pdf Accessed June 25, 2024.

BCR-ABL1 Testing in Chronic Myelogenous Leukemia and Acute Lymphoblastic Leukemia

- 1. Jabbour E, Kantarjian H. Chronic myeloid leukemia: 2022 update on diagnosis, therapy, and monitoring. Am J Hematol. Sep 2022; 97(9): 1236-1256. PMID 35751859
- Sawyers CL. Chronic myeloid leukemia. N Engl J Med. Apr 29 1999; 340(17): 1330-40.
 PMID 10219069
- 3. Kantarjian HM, Deisseroth A, Kurzrock R, et al. Chronic myelogenous leukemia: a concise update. Blood. Aug 01 1993; 82(3): 691-703. PMID 8338938
- 4. Savage DG, Szydlo RM, Chase A, et al. Bone marrow transplantation for chronic myeloid leukaemia: the effects of differing criteria for defining chronic phase on probabilities of survival and relapse. Br J Haematol. Oct 1997; 99(1): 30-5. PMID 9359498
- 5. Arber DA, Orazi A, Hasserjian RP, et al. International Consensus Classification of Myeloid Neoplasms and Acute Leukemias: integrating morphologic, clinical, and genomic data. Blood. Sep 15 2022; 140(11): 1200-1228. PMID 35767897
- 6. Khoury JD, Solary E, Abla O, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Myeloid and Histiocytic/Dendritic Neoplasms. Leukemia. Jul 2022; 36(7): 1703-1719. PMID 35732831
- 7. Malard F, Mohty M. Acute lymphoblastic leukaemia. Lancet. Apr 04 2020; 395(10230): 1146-1162. PMID 32247396
- 8. Esparza SD, Sakamoto KM. Topics in pediatric leukemia--acute lymphoblastic leukemia. MedGenMed. Mar 07 2005; 7(1): 23. PMID 16369328
- 9. Jabbour EJ, Faderl S, Kantarjian HM. Adult acute lymphoblastic leukemia. Mayo Clin Proc. Nov 2005; 80(11): 1517-27. PMID 16295033
- 10. National Cancer Institute Surveillance, Epidemiology, and End Results Program. Cancer Stat Facts: Leukemia Acute Lymphocytic Leukemia (ALL). 2020. https://seer.cancer.gov/statfacts/html/alyl.html. Accessed August 13, 2024.
- 11. Mullighan CG. The molecular genetic makeup of acute lymphoblastic leukemia. Hematology Am Soc Hematol Educ Program. 2012; 2012: 389-96. PMID 23233609
- 12. Hughes T, Deininger M, Hochhaus A, et al. Monitoring CML patients responding to treatment with tyrosine kinase inhibitors: review and recommendations for harmonizing current methodology for detecting BCR-ABL transcripts and kinase domain mutations and for expressing results. Blood. Jul 01 2006; 108(1): 28-37. PMID 16522812

- 13. Cross NC. Standardisation of molecular monitoring for chronic myeloid leukaemia. Best Pract Res Clin Haematol. Sep 2009; 22(3): 355-65. PMID 19959086
- 14. Hughes T, Branford S. Molecular monitoring of BCR-ABL as a guide to clinical management in chronic myeloid leukaemia. Blood Rev. Jan 2006; 20(1): 29-41. PMID 16426942
- National Comprehensive Cancer Network (NCCN). NCCN clinical practice guidelines in oncology: Acute Lymphoblastic Leukemia. Version 2.2024. https://www.nccn.org/professionals/physician_gls/pdf/all.pdf. Accessed August 13, 2024.
- 16. Jones D, Kamel-Reid S, Bahler D, et al. Laboratory practice guidelines for detecting and reporting BCR-ABL drug resistance mutations in chronic myelogenous leukemia and acute lymphoblastic leukemia: a report of the Association for Molecular Pathology. J Mol Diagn. Jan 2009; 11(1): 4-11. PMID 19095773
- 17. Saglio G, Kim DW, Issaragrisil S, et al. Nilotinib versus imatinib for newly diagnosed chronic myeloid leukemia. N Engl J Med. Jun 17 2010; 362(24): 2251-9. PMID 20525993
- 18. Kantarjian H, Shah NP, Hochhaus A, et al. Dasatinib versus imatinib in newly diagnosed chronic-phase chronic myeloid leukemia. N Engl J Med. Jun 17 2010; 362(24): 2260-70. PMID 20525995
- 19. Cortes JE, Gambacorti-Passerini C, Deininger MW, et al. Bosutinib Versus Imatinib for Newly Diagnosed Chronic Myeloid Leukemia: Results From the Randomized BFORE Trial. J Clin Oncol. Jan 20 2018; 36(3): 231-237. PMID 29091516
- National Comprehensive Cancer Network (NCCN). NCCN clinical practice guidelines in oncology: Chronic Myeloid Leukemia. Version 1.2025. https://www.nccn.org/professionals/physician_gls/pdf/cml.pdf. Accessed August 12, 2024.
- 21. Mughal TI, Goldman JM. Emerging strategies for the treatment of mutant Bcr-Abl T315I myeloid leukemia. Clin Lymphoma Myeloma. Mar 2007; 7 Suppl 2: S81-4. PMID 17382017
- 22. von Bubnoff N, Manley PW, Mestan J, et al. Bcr-Abl resistance screening predicts a limited spectrum of point mutations to be associated with clinical resistance to the Abl kinase inhibitor nilotinib (AMN107). Blood. Aug 15 2006; 108(4): 1328-33. PMID 16614241
- 23. Piccaluga PP, Martinelli G, Rondoni M, et al. Advances and potential treatment for Philadelphia chromosome-positive adult acute lymphoid leukaemia. Expert Opin Biol Ther. Oct 2006; 6(10): 1011-22. PMID 16989583
- 24. Guilhot F, Apperley J, Kim DW, et al. Dasatinib induces significant hematologic and cytogenetic responses in patients with imatinib-resistant or -intolerant chronic myeloid leukemia in accelerated phase. Blood. May 15 2007; 109(10): 4143-50. PMID 17264298
- 25. Cortes J, Rousselot P, Kim DW, et al. Dasatinib induces complete hematologic and cytogenetic responses in patients with imatinib-resistant or -intolerant chronic myeloid leukemia in blast crisis. Blood. Apr 15 2007; 109(8): 3207-13. PMID 17185463
- 26. Walz C, Sattler M. Novel targeted therapies to overcome imatinib mesylate resistance in chronic myeloid leukemia (CML). Crit Rev Oncol Hematol. Feb 2006; 57(2): 145-64. PMID 16213151
- 27. Cortes J, Kantarjian H. How I treat newly diagnosed chronic phase CML. Blood. Aug 16 2012; 120(7): 1390-7. PMID 22613793

- 28. Branford S, Hughes TP, Rudzki Z. Monitoring chronic myeloid leukaemia therapy by real-time quantitative PCR in blood is a reliable alternative to bone marrow cytogenetics. Br J Haematol. Dec 1999; 107(3): 587-99. PMID 10583264
- 29. Radich JP. Measuring response to BCR-ABL inhibitors in chronic myeloid leukemia. Cancer. Jan 15 2012; 118(2): 300-11. PMID 21717440
- 30. Campiotti L, Suter MB, Guasti L, et al. Imatinib discontinuation in chronic myeloid leukaemia patients with undetectable BCR-ABL transcript level: A systematic review and a meta-analysis. Eur J Cancer. May 2017; 77: 48-56. PMID 28365527
- 31. Druker BJ, Guilhot F, O'Brien SG, et al. Five-year follow-up of patients receiving imatinib for chronic myeloid leukemia. N Engl J Med. Dec 07 2006; 355(23): 2408-17. PMID 17151364
- 32. Boeckx N, Laer CV, Roover JD, et al. Comparison of molecular responses based on BCR-ABL1% (IS) results from an in-house TaqMan-based qPCR versus Xpert(®) assay in CML patients on tyrosine kinase inhibitor therapy. Acta Clin Belg. Aug 2015; 70(4): 237-43. PMID 26166681
- 33. Etienne G, Guilhot J, Rea D, et al. Long-Term Follow-Up of the French Stop Imatinib (STIM1) Study in Patients With Chronic Myeloid Leukemia. J Clin Oncol. Jan 20 2017; 35(3): 298-305. PMID 28095277
- 34. Clark RE, Polydoros F, Apperley JF, et al. De-escalation of tyrosine kinase inhibitor therapy before complete treatment discontinuation in patients with chronic myeloid leukaemia (DESTINY): a non-randomised, phase 2 trial. Lancet Haematol. Jul 2019; 6(7): e375-e383. PMID 31201085
- 35. Devos T, Verhoef G, Steel E, et al. Interruption or Discontinuation of Tyrosine Kinase Inhibitor Treatment in Chronic Myeloid Leukaemia: A Retrospective Cohort Study (SPARKLE) in Belgium. Acta Haematol. 2019; 142(4): 197-207. PMID 31163431
- 36. Saussele S, Richter J, Guilhot J, et al. Discontinuation of tyrosine kinase inhibitor therapy in chronic myeloid leukaemia (EURO-SKI): a prespecified interim analysis of a prospective, multicentre, non-randomised, trial. Lancet Oncol. Jun 2018; 19(6): 747-757. PMID 29735299
- 37. Ross DM, Masszi T, Gómez Casares MT, et al. Durable treatment-free remission in patients with chronic myeloid leukemia in chronic phase following frontline nilotinib: 96-week update of the ENESTfreedom study. J Cancer Res Clin Oncol. May 2018; 144(5): 945-954. PMID 29468438
- 38. Wang R, Cong Y, Li C, et al. Predictive value of early molecular response for deep molecular response in chronic phase of chronic myeloid leukemia. Medicine (Baltimore). Apr 2019; 98(15): e15222. PMID 30985724
- 39. Berdeja JG, Heinrich MC, Dakhil SR, et al. Rates of deep molecular response by digital and conventional PCR with frontline nilotinib in newly diagnosed chronic myeloid leukemia: a landmark analysis. Leuk Lymphoma. Oct 2019; 60(10): 2384-2393. PMID 30912699
- 40. Shah NP, García-Gutiérrez V, Jiménez-Velasco A, et al. Dasatinib discontinuation in patients with chronic-phase chronic myeloid leukemia and stable deep molecular response: the DASFREE study. Leuk Lymphoma. Mar 2020; 61(3): 650-659. PMID 31647335
- 41. Shah NP, García-Gutiérrez V, Jiménez-Velasco A, et al. Treatment-free remission after dasatinib in patients with chronic myeloid leukaemia in chronic phase with deep

- molecular response: Final 5-year analysis of DASFREE. Br J Haematol. Sep 2023; 202(5): 942-952. PMID 37246588
- 42. Press RD, Love Z, Tronnes AA, et al. BCR-ABL mRNA levels at and after the time of a complete cytogenetic response (CCR) predict the duration of CCR in imatinib mesylate-treated patients with CML. Blood. Jun 01 2006; 107(11): 4250-6. PMID 16467199
- 43. Branford S, Rudzki Z, Harper A, et al. Imatinib produces significantly superior molecular responses compared to interferon alfa plus cytarabine in patients with newly diagnosed chronic myeloid leukemia in chronic phase. Leukemia. Dec 2003; 17(12): 2401-9. PMID 14523461
- 44. Nicolini FE, Dulucq S, Boureau L, et al. Evaluation of Residual Disease and TKI Duration Are Critical Predictive Factors for Molecular Recurrence after Stopping Imatinib First-line in Chronic Phase CML Patients. Clin Cancer Res. Nov 15 2019; 25(22): 6606-6613. PMID 31292142
- 45. Yan D, Pomicter AD, O'Hare T, et al. ddeeper Than Deep: Can ddPCR Predict Successful Imatinib Cessation?. Clin Cancer Res. Nov 15 2019; 25(22): 6561-6563. PMID 31540978
- 46. Atallah E, Schiffer CA, Radich JP, et al. Assessment of Outcomes After Stopping Tyrosine Kinase Inhibitors Among Patients With Chronic Myeloid Leukemia: A Nonrandomized Clinical Trial. JAMA Oncol. Jan 01 2021; 7(1): 42-50. PMID 33180106
- 47. Haddad FG, Sasaki K, Issa GC, et al. Treatment-free remission in patients with chronic myeloid leukemia following the discontinuation of tyrosine kinase inhibitors. Am J Hematol. Jul 2022; 97(7): 856-864. PMID 35357036
- 48. Hehlmann R, Lauseker M, Jung-Munkwitz S, et al. Tolerability-adapted imatinib 800 mg/d versus 400 mg/d versus 400 mg/d plus interferon-α in newly diagnosed chronic myeloid leukemia. J Clin Oncol. Apr 20 2011; 29(12): 1634-42. PMID 21422420
- 49. Wang L, Pearson K, Ferguson JE, et al. The early molecular response to imatinib predicts cytogenetic and clinical outcome in chronic myeloid leukaemia. Br J Haematol. Mar 2003; 120(6): 990-9. PMID 12648069
- 50. Quintás-Cardama A, Kantarjian H, Jones D, et al. Delayed achievement of cytogenetic and molecular response is associated with increased risk of progression among patients with chronic myeloid leukemia in early chronic phase receiving high-dose or standard-dose imatinib therapy. Blood. Jun 18 2009; 113(25): 6315-21. PMID 19369233
- 51. Campana D. Should minimal residual disease monitoring in acute lymphoblastic leukemia be standard of care?. Curr Hematol Malig Rep. Jun 2012; 7(2): 170-7. PMID 22373809
- 52. Muller MC, Hanfstein B, Erben P, et al. Molecular response to first line imatinib therapy is predictive for long term event free survival in patients with chronic phase chronic myelogenous leukemia: an interim analysis of the randomized German CML Study IV. Blood 2008;112:129. Abstract 333.
- 53. Press RD, Galderisi C, Yang R, et al. A half-log increase in BCR-ABL RNA predicts a higher risk of relapse in patients with chronic myeloid leukemia with an imatinib-induced complete cytogenetic response. Clin Cancer Res. Oct 15 2007; 13(20): 6136-43. PMID 17947479
- 54. Marin D, Milojkovic D, Olavarria E, et al. European LeukemiaNet criteria for failure or suboptimal response reliably identify patients with CML in early chronic phase treated

- with imatinib whose eventual outcome is poor. Blood. Dec 01 2008; 112(12): 4437-44. PMID 18716134
- 55. Baccarani M, Castagnetti F, Gugliotta G, et al. A review of the European LeukemiaNet recommendations for the management of CML. Ann Hematol. Apr 2015; 94 Suppl 2: \$141-7. PMID 25814080
- 56. Branford S, Rudzki Z, Parkinson I, et al. Real-time quantitative PCR analysis can be used as a primary screen to identify patients with CML treated with imatinib who have BCR-ABL kinase domain mutations. Blood. Nov 01 2004; 104(9): 2926-32. PMID 15256429
- 57. Wang L, Knight K, Lucas C, et al. The role of serial BCR-ABL transcript monitoring in predicting the emergence of BCR-ABL kinase mutations in imatinib-treated patients with chronic myeloid leukemia. Haematologica. Feb 2006; 91(2): 235-9. PMID 16461309
- 58. Soverini S, Hochhaus A, Nicolini FE, et al. BCR-ABL kinase domain mutation analysis in chronic myeloid leukemia patients treated with tyrosine kinase inhibitors: recommendations from an expert panel on behalf of European LeukemiaNet. Blood. Aug 04 2011; 118(5): 1208-15. PMID 21562040
- 59. Terasawa T, Dahabreh I, Castaldi PJ, et al. Systematic reviews on selected pharmacogenetic tests for cancer treatment: CYP2D6 for Tamoxifen in breast cancer, KRAS for anti-EGFR antibodies in colorectal cancer, and BCR-ABL1 for tyrosine kinase inhibitors in chronic myeloid leukemia. Rockville, MD: Agency for Healthcare Research and Quality; 2010.
- 60. Xue M, Cheng J, Zhao J, et al. Outcomes of 219 chronic myeloid leukaemia patients with additional chromosomal abnormalities and/or tyrosine kinase domain mutations. Int J Lab Hematol. Feb 2019; 41(1): 94-101. PMID 30285321
- 61. Branford S, Melo JV, Hughes TP. Selecting optimal second-line tyrosine kinase inhibitor therapy for chronic myeloid leukemia patients after imatinib failure: does the BCR-ABL mutation status really matter?. Blood. Dec 24 2009; 114(27): 5426-35. PMID 19880502
- 62. Cortes JE, Kim DW, Pinilla-Ibarz J, et al. A Pivotal Phase 2 Trial of Ponatinib in Patients with Chronic Myeloid Leukemia (CML) and Philadelphia Chromosome-Positive Acute Lymphoblastic Leukemia (Ph+ALL) Resistant or Intolerant to Dasatinib or Nilotinib, or with the T315I BCR-ABL Mutation: 12-Month Follow-up of the PACE Trial. American Society of Hematology 54th Annual Meeting, December 2012. 2012:Abstract 163.
- 63. Ernst T, Gruber FX, Pelz-Ackermann O, et al. A co-operative evaluation of different methods of detecting BCR-ABL kinase domain mutations in patients with chronic myeloid leukemia on second-line dasatinib or nilotinib therapy after failure of imatinib. Haematologica. Sep 2009; 94(9): 1227-35. PMID 19608684
- 64. Alikian M, Gerrard G, Subramanian PG, et al. BCR-ABL1 kinase domain mutations: methodology and clinical evaluation. Am J Hematol. Mar 2012; 87(3): 298-304. PMID 22231203
- 65. Fielding AK, Zakout GA. Treatment of Philadelphia chromosome-positive acute lymphoblastic leukemia. Curr Hematol Malig Rep. Jun 2013; 8(2): 98-108. PMID 23475624
- 66. Campana D. Minimal residual disease in acute lymphoblastic leukemia. Hematology Am Soc Hematol Educ Program. 2010; 2010: 7-12. PMID 21239764

- 67. Arunachalam AK, Janet NB, Korula A, et al. Prognostic value of MRD monitoring based on BCR-ABL1 copy numbers in Philadelphia chromosome positive acute lymphoblastic leukemia. Leuk Lymphoma. Dec 2020; 61(14): 3468-3475. PMID 32852239
- 68. Conter V, Bartram CR, Valsecchi MG, et al. Molecular response to treatment redefines all prognostic factors in children and adolescents with B-cell precursor acute lymphoblastic leukemia: results in 3184 patients of the AIEOP-BFM ALL 2000 study. Blood. Apr 22 2010; 115(16): 3206-14. PMID 20154213
- 69. Soverini S, De Benedittis C, Polakova KM, et al. Next-generation sequencing for sensitive detection of BCR-ABL1 mutations relevant to tyrosine kinase inhibitor choice in imatinib-resistant patients. Oncotarget. Apr 19 2016; 7(16): 21982-90. PMID 26980736

<u>Carrier Screening for Genetic Diseases</u>

- 1. Henneman L, Borry P, Chokoshvili D, et al. Responsible implementation of expanded carrier screening. Eur J Hum Genet. Jun 2016; 24(6): e1-e12. PMID 26980105
- 2. Edwards JG, Feldman G, Goldberg J, et al. Expanded carrier screening in reproductive medicine-points to consider: a joint statement of the American College of Medical Genetics and Genomics, American College of Obstetricians and Gynecologists, National Society of Genetic Counselors, Perinatal Quality Foundation, and Society for Maternal-Fetal Medicine. Obstet Gynecol. Mar 2015; 125(3): 653-662. PMID 25730230
- 3. Kaback MM. Population-based genetic screening for reproductive counseling: the Tay-Sachs disease model. Eur J Pediatr. Dec 2000; 159 Suppl 3: S192-5. PMID 11216898
- 4. Banda Y, Kvale MN, Hoffmann TJ, et al. Characterizing Race/Ethnicity and Genetic Ancestry for 100,000 Subjects in the Genetic Epidemiology Research on Adult Health and Aging (GERA) Cohort. Genetics. Aug 2015; 200(4): 1285-95. PMID 26092716
- 5. Grant MD, Lauderdale DS. Cohort effects in a genetically determined trait: eye colour among US whites. Ann Hum Biol. 2002; 29(6): 657-66. PMID 12573082
- 6. Committee on Genetics, American College of Obstetricians and Gynecologists. ACOG Committee Opinion. Number 325, December 2005. Update on carrier screening for cystic fibrosis. Obstet Gynecol. Dec 2005; 106(6): 1465-8. PMID 16319281
- 7. Committee Opinion No. 691: Carrier Screening for Genetic Conditions. Obstet Gynecol. Mar 2017; 129(3): e41-e55. PMID 28225426
- 8. Watson MS, Cutting GR, Desnick RJ, et al. Cystic fibrosis population carrier screening: 2004 revision of American College of Medical Genetics mutation panel. Genet Med. 2004; 6(5): 387-91. PMID 15371902
- 9. Deignan JL, Gregg AR, Grody WW, et al. Updated recommendations for CFTR carrier screening: A position statement of the American College of Medical Genetics and Genomics (ACMG). Genet Med. Aug 2023; 25(8): 100867. PMID 37310422
- 10. Prior TW. Carrier screening for spinal muscular atrophy. Genet Med. Nov 2008; 10(11): 840-2. PMID 18941424
- 11. Gross SJ, Pletcher BA, Monaghan KG. Carrier screening in individuals of Ashkenazi Jewish descent. Genet Med. Jan 2008; 10(1): 54-6. PMID 18197057
- 12. Burke W, Tarini B, Press NA, et al. Genetic screening. Epidemiol Rev. 2011; 33(1): 148-64. PMID 21709145
- 13. Bajaj K, Gross SJ. Carrier screening: past, present, and future. J Clin Med. Sept 2015 2014;3(3):1033-1042. PMC4449659

- 14. Kaback M, Lim-Steele J, Dabholkar D, et al. Tay-Sachs disease--carrier screening, prenatal diagnosis, and the molecular era. An international perspective, 1970 to 1993. The International TSD Data Collection Network. JAMA. Nov 17 1993; 270(19): 2307-15. PMID 8230592
- 15. Ioannou L, McClaren BJ, Massie J, et al. Population-based carrier screening for cystic fibrosis: a systematic review of 23 years of research. Genet Med. Mar 2014; 16(3): 207-16. PMID 24030436
- 16. Wang T, Kiss D, McFadden K, et al. Clinical utility of reproductive carrier screening for preconception and pregnant couples for multiple genetic conditions: a systematic review and meta-analysis. Expert Rev Mol Diagn. May 2023; 23(5): 419-429. PMID 37086152
- 17. Wang M, Schaink A, Holubowich C, et al. Carrier Screening Programs for Cystic Fibrosis, Fragile X Syndrome, Hemoglobinopathies and Thalassemia, and Spinal Muscular Atrophy: A Health Technology Assessment. Ont Health Technol Assess Ser. 2023; 23(4): 1-398. PMID 37637488
- 18. Shraga R, Yarnall S, Elango S, et al. Evaluating genetic ancestry and self-reported ethnicity in the context of carrier screening. BMC Genet. Nov 28 2017; 18(1): 99. PMID 29179688
- 19. Committee Opinion No. 690: Carrier Screening in the Age of Genomic Medicine. Obstet Gynecol. Mar 2017; 129(3): e35-e40. PMID 28225425
- 20. Gregg AR, Aarabi M, Klugman S, et al. Screening for autosomal recessive and X-linked conditions during pregnancy and preconception: a practice resource of the American College of Medical Genetics and Genomics (ACMG). Genet Med. Oct 2021; 23(10): 1793-1806. PMID 34285390
- 21. Stevens B, Krstic N, Jones M, et al. Finding Middle Ground in Constructing a Clinically Useful Expanded Carrier Screening Panel. Obstet Gynecol. Aug 2017; 130(2): 279-284. PMID 28697118
- 22. Kaseniit KE, Haque IS, Goldberg JD, et al. Genetic ancestry analysis on 93,000 individuals undergoing expanded carrier screening reveals limitations of ethnicity-based medical guidelines. Genet Med. Oct 2020; 22(10): 1694-1702. PMID 32595206
- 23. Westemeyer M, Saucier J, Wallace J, et al. Clinical experience with carrier screening in a general population: support for a comprehensive pan-ethnic approach. Genet Med. Aug 2020; 22(8): 1320-1328. PMID 32366966
- 24. Guo MH, Gregg AR. Estimating yields of prenatal carrier screening and implications for design of expanded carrier screening panels. Genet Med. Sep 2019; 21(9): 1940-1947. PMID 30846881
- 25. Terhaar C, Teed N, Allen R, et al. Clinical experience with multigene carrier panels in the reproductive setting. Prenat Diagn. Apr 23 2018; 38(8): 572-7. PMID 29683194
- 26. Peyser A, Singer T, Mullin C, et al. Comparing ethnicity-based and expanded carrier screening methods at a single fertility center reveals significant differences in carrier rates and carrier couple rates. Genet Med. Jun 2019; 21(6): 1400-1406. PMID 30327537
- 27. Hernandez-Nieto C, Alkon-Meadows T, Lee J, et al. Expanded carrier screening for preconception reproductive risk assessment: Prevalence of carrier status in a Mexican population. Prenat Diagn. Apr 2020; 40(5): 635-643. PMID 32003480
- 28. Ben-Shachar R, Svenson A, Goldberg JD, et al. A data-driven evaluation of the size and content of expanded carrier screening panels. Genet Med. Sep 2019; 21(9): 1931-1939. PMID 30816298

- 29. Arjunan A, Bellerose H, Torres R, et al. Evaluation and classification of severity for 176 genes on an expanded carrier screening panel. Prenat Diagn. Sep 2020; 40(10): 1246-1257. PMID 32474937
- 30. Kirk EP, Delatycki MB, Archibald AD, et al. Nationwide, Couple-Based Genetic Carrier Screening. N Engl J Med. Nov 21 2024; 391(20): 1877-1889. PMID 39565987
- 31. Beauchamp KA, Johansen Taber KA, Muzzey D. Clinical impact and cost-effectiveness of a 176-condition expanded carrier screen. Genet Med. Sep 2019; 21(9): 1948-1957. PMID 30760891
- 32. Ghiossi CE, Goldberg JD, Haque IS, et al. Clinical Utility of Expanded Carrier Screening: Reproductive Behaviors of At-Risk Couples. J Genet Couns. Jun 2018; 27(3): 616-625. PMID 28956228
- 33. Johansen Taber KA, Beauchamp KA, Lazarin GA, et al. Clinical utility of expanded carrier screening: results-guided actionability and outcomes. Genet Med. May 2019; 21(5): 1041-1048. PMID 30310157

Related Policies

Assays of Genetic Expression in Tumor Tissue as a Technique to Determine Prognosis in Patients with Breast Cancer

Document Precedence

Blue Cross and Blue Shield of Vermont (Blue Cross VT) Medical Policies are developed to provide clinical guidance and are based on research of current medical literature and review of common medical practices in the treatment and diagnosis of disease. The applicable group/individual contract and member certificate language, or employer's benefit plan if an ASO group, determines benefits that are in effect at the time of service. Since medical practices and knowledge are constantly evolving, Blue Cross VT reserves the right to review and revise its medical policies periodically. To the extent that there may be any conflict between medical policy and contract/employer benefit plan language, the member's contract/employer benefit plan language takes precedence.

Audit Information

Blue Cross VT reserves the right to conduct audits on any provider and/or facility to ensure compliance with the guidelines stated in the medical policy. If an audit identifies instances of non-compliance with this medical policy, Blue Cross VT reserves the right to recoup all non-compliant payments.

Administrative and Contractual Guidance

Benefit Determination Guidance

Prior approval may be required for services outlined in this policy. Benefits are subject to all terms, limitations and conditions of the subscriber contract.

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Incomplete authorization requests may result in a delay of decision pending submission of missing information. To be considered complete, see policy guidelines above.

NEHP/ABNE members may have different benefits for services listed in this policy. To confirm benefits, please contact the customer service department at the member's health plan.

Federal Employee Program (FEP): Members may have different benefits that apply. For further information please contact FEP customer service or refer to the FEP Service Benefit Plan Brochure. It is important to verify the member's benefits prior to providing the service to determine if benefits are available or if there is a specific exclusion in the member's benefit.

Coverage varies according to the member's group or individual contract. Not all groups are required to follow the Vermont legislative mandates. Member Contract language takes precedence over medical policy when there is a conflict.

If the member receives benefits through an Administrative Services Only (ASO) group, benefits may vary or not apply. To verify benefit information, please refer to the member's employer benefit plan documents or contact the customer service department. Language in the employer benefit plan documents takes precedence over medical policy when there is a conflict.

Policy Implementation/Update information

08/11/2025	New Policy

Eligible providers

Qualified healthcare professionals practicing within the scope of their license(s).

Approved by Blue Cross VT Medical Directors

Tom Weigel, MD, MBA Vice President and Chief Medical Officer

Tammaji P. Kulkarni, MD Senior Medical Director

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Attachment I - Coding Table

Code Type	Number	Description	Policy Instructions			
	The following codes will be considered as medically necessary when applicable criteria have been met.					
CPT®	81162	BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full sequence analysis and full duplication/deletion analysis (ie, detection of large gene rearrangements)	Requires Prior Approval			
CPT®	81163	BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full sequence analysis	Requires Prior Approval			
CPT®	81164	BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full duplication/deletion analysis (ie, detection of large gene rearrangements)	Requires Prior Approval			
CPT®	81165	BRCA1 (BRCA1, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full sequence analysis	Requires Prior Approval			
CPT®	81166	BRCA1 (BRCA1, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full duplication/deletion analysis (ie, detection of large gene rearrangements)	Requires Prior Approval			
CPT®	81167	BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full duplication/deletion analysis (ie, detection of large gene rearrangements)	Requires Prior Approval			
CPT®	81170	ABL1 (ABL proto-oncogene 1, non- receptor tyrosine kinase) (eg, acquired imatinib tyrosine kinase inhibitor resistance), gene analysis, variants in the kinase domain	Requires Prior Approval			
CPT®	81201	APC (adenomatous polyposis coli) (eg, familial adenomatosis polyposis [FAP],	Requires Prior Approval			

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Code Type	Number	Description	Policy Instructions
		attenuated FAP) gene analysis; full gene sequence	
CPT®	81202	APC (adenomatous polyposis coli) (eg, familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; known familial variants	Requires Prior Approval
CPT®	81203	APC (adenomatous polyposis coli) (eg, familial adenomatosis polyposis [FAP], attenuated FAP) gene analysis; duplication/deletion variants	Requires Prior Approval
CPT®	81206	BCR/ABL1 (t(9;22)) (eg, chronic myelogenous leukemia) translocation analysis; major breakpoint, qualitative or quantitative	Requires Prior Approval
CPT®	81207	BCR/ABL1 (t(9;22)) (eg, chronic myelogenous leukemia) translocation analysis; minor breakpoint, qualitative or quantitative	Requires Prior Approval
CPT®	81208	BCR/ABL1 (t(9;22)) (eg, chronic myelogenous leukemia) translocation analysis; other breakpoint, qualitative or quantitative	Requires Prior Approval
CPT®	81210	BRAF (B-Raf proto-oncogene, serine/threonine kinase) (eg, colon cancer, melanoma), gene analysis, V600 variant(s)	Requires Prior Approval
CPT®	81212	BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; 185delAG, 5385insC, 6174delT variants	Requires Prior Approval
CPT®	81215	BRCA1 (BRCA1, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; known familial variant	Requires Prior Approval
CPT®	81216	BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; full sequence analysis	Requires Prior Approval
CPT®	81217	BRCA2 (BRCA2, DNA repair associated) (eg, hereditary breast and ovarian cancer) gene analysis; known familial variant	Requires Prior Approval

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Code Type	Number	Description	Policy Instructions
CPT®	81219	CALR (calreticulin) (eg, myeloproliferative disorders), gene analysis, common variants in exon 9	Requires Prior Approval
CPT®	81270	JAK2 (Janus kinase 2) (eg, myeloproliferative disorder) gene analysis, p.Val617Phe (V617F) variant	Requires Prior Approval
CPT®	81279	JAK2 (Janus kinase 2) (eg, myeloproliferative disorder) targeted sequence analysis (eg, exons 12 and 13)	Requires Prior Approval
CPT®	81288	MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; promoter methylation analysis	Requires Prior Approval
CPT®	81292	MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis	Requires Prior Approval
CPT®	81293	MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants	Requires Prior Approval
CPT®	81294	MLH1 (mutL homolog 1, colon cancer, nonpolyposis type 2) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants	Requires Prior Approval
CPT®	81295	MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis	Requires Prior Approval
CPT®	81296	MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants	Requires Prior Approval
CPT®	81297	MSH2 (mutS homolog 2, colon cancer, nonpolyposis type 1) (eg, hereditary nonpolyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants	Requires Prior Approval

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Code Type	Number	Description	Policy Instructions
CPT®	81298	MSH6 (mutS homolog 6 [E. coli]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis	Requires Prior Approval
CPT®	81299	MSH6 (mutS homolog 6 [E. coli]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants	Requires Prior Approval
CPT®	81300	MSH6 (mutS homolog 6 [E. coli]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants	Requires Prior Approval
CPT®	81301	Microsatellite instability analysis (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) of markers for mismatch repair deficiency (eg, BAT25, BAT26), includes comparison of neoplastic and normal tissue, if performed	Requires Prior Approval
CPT®	81307	PALB2 (partner and localizer of BRCA2) (eg, breast and pancreatic cancer) gene analysis; full gene sequence	Requires Prior Approval
CPT®	81308	PALB2 (partner and localizer of BRCA2) (eg, breast and pancreatic cancer) gene analysis; known familial variant	Requires Prior Approval
CPT®	81317	PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; full sequence analysis	Requires Prior Approval
CPT®	81318	PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; known familial variants	Requires Prior Approval
CPT®	81319	PMS2 (postmeiotic segregation increased 2 [S. cerevisiae]) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) gene analysis; duplication/deletion variants	Requires Prior Approval
CPT®	81338	MPL (MPL proto-oncogene, thrombopoietin receptor) (eg, myeloproliferative disorder) gene	Requires Prior Approval

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Code Type	Number	Description	Policy Instructions
		analysis; common variants (eg, W515A, W515K, W515L, W515R)	
CPT®	81339	MPL (MPL proto-oncogene, thrombopoietin receptor) (eg, myeloproliferative disorder) gene analysis; sequence analysis, exon 10	Requires Prior Approval
CPT®	81401	Molecular pathology procedure, Level 2 (eg, 2-10 SNPs, 1 methylated variant, or 1 somatic variant [typically using nonsequencing target variant analysis], or detection of a dynamic mutation disorder/triplet repeat)	Requires Prior Approval
CPT®	81403	Molecular pathology procedure, Level 4 (eg, analysis of single exon by DNA sequence analysis, analysis of >10 amplicons using multiplex PCR in 2 or more independent reactions, mutation scanning or duplication/deletion variants of 2-5 exons)	Requires Prior Approval
CPT®	81412	Ashkenazi Jewish associated disorders (eg, Bloom syndrome, Canavan disease, cystic fibrosis, familial dysautonomia, Fanconi anemia group C, Gaucher disease, Tay-Sachs disease), genomic sequence analysis panel, must include sequencing of at least 9 genes, including ASPA, BLM, CFTR, FANCC, GBA, HEXA, IKBKAP, MCOLN1, and SMPD1	Requires Prior Approval
CPT®	81432	Hereditary breast cancer-related disorders (eg, hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer, hereditary pancreatic cancer, hereditary prostate cancer), genomic sequence analysis panel, 5 or more genes, interrogation for sequence variants and copy number variants	Requires Prior Approval
CPT®	81435	Hereditary colon cancer-related disorders (eg, Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), genomic sequence analysis panel, 5 or more genes, interrogation for sequence variants and copy number variants	Requires Prior Approval

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Code Type	Number	Description	Policy Instructions
CPT®	81443	Genetic testing for severe inherited conditions (eg, cystic fibrosis, Ashkenazi Jewish-associated disorders [eg, Bloom syndrome, Canavan disease, Fanconi anemia type C, mucolipidosis type VI, Gaucher disease, Tay-Sachs disease], beta hemoglobinopathies, phenylketonuria, galactosemia), genomic sequence analysis panel, must include sequencing of at least 15 genes (eg, ACADM, ARSA, ASPA, ATP7B, BCKDHA, BCKDHB, BLM, CFTR, DHCR7, FANCC, G6PC, GAA, GALT, GBA, GBE1, HBB, HEXA, IKBKAP, MCOLN1, PAH)	Requires Prior Approval
CPT®	81479	Unlisted molecular pathology procedure	Suspend for Medical Review
CPT®	0016U	Oncology (hematolymphoid neoplasia), RNA, BCR/ABL1 major and minor breakpoint fusion transcripts, quantitative PCR amplification, blood or bone marrow, report of fusion not detected or detected with quantitation	No Prior Approval Required
CPT®	0017U	Oncology (hematolymphoid neoplasia), JAK2 mutation, DNA, PCR amplification of exons 12-14 and sequence analysis, blood or bone marrow, report of JAK2 mutation not detected or detected	No Prior Approval Required
CPT®	0027U	JAK2 (Janus kinase 2) (eg, myeloproliferative disorder) gene analysis, targeted sequence analysis exons 12-15	No Prior Approval Required
CPT®	0040U	BCR/ABL1 (t(9;22)) (eg, chronic myelogenous leukemia) translocation analysis, major breakpoint, quantitative	No Prior Approval Required
CPT®	0129U	Hereditary breast cancer-related disorders (eg, hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer), genomic sequence analysis and deletion/duplication analysis panel (ATM, BRCA1, BRCA2, CDH1, CHEK2, PALB2, PTEN, and TP53)	Requires Prior Approval
CPT®	0157U	APC (APC regulator of WNT signaling pathway) (eg, familial adenomatosis polyposis [FAP]) mRNA sequence analysis	Requires Prior Approval

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Code Type	Number	Description	Policy Instructions
		(List separately in addition to code for primary procedure)	
CPT®	0158U	MLH1 (mutL homolog 1) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)	Requires Prior Approval
CPT®	0159U	MSH2 (mutS homolog 2) (eg, hereditary colon cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)	Requires Prior Approval
CPT®	0160U	MSH6 (mutS homolog 6) (eg, hereditary colon cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)	Requires Prior Approval
CPT®	0161U	PMS2 (PMS1 homolog 2, mismatch repair system component) (eg, hereditary non-polyposis colorectal cancer, Lynch syndrome) mRNA sequence analysis (List separately in addition to code for primary procedure)	Requires Prior Approval
CPT®	0162U	Hereditary colon cancer (Lynch syndrome), targeted mRNA sequence analysis panel (MLH1, MSH2, MSH6, PMS2) (List separately in addition to code for primary procedure)	Requires Prior Approval
CPT®	0172U	Oncology (solid tumor as indicated by the label), somatic mutation analysis of BRCA1 (BRCA1, DNA repair associated), BRCA2 (BRCA2, DNA repair associated) and analysis of homologous recombination deficiency pathways, DNA, formalin-fixed paraffin-embedded tissue, algorithm quantifying tumor genomic instability score	Requires Prior Approval
CPT®	0238U	Oncology (Lynch syndrome), genomic DNA sequence analysis of MLH1, MSH2, MSH6, PMS2, and EPCAM, including small sequence changes in exonic and intronic regions, deletions, duplications, mobile element insertions, and variants in non-uniquely mappable regions	Requires Prior Approval
The following codes will be denied and Not Medically Necessary, Non-Covered, Contract Exclusions or Investigational.			

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Code Type	Number	Description	Policy Instructions
CPT®	0101U	Hereditary colon cancer disorders (eg, Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), genomic sequence analysis panel utilizing a combination of NGS, Sanger, MLPA, and array CGH, with mRNA analytics to resolve variants of unknown significance when indicated (15 genes [sequencing and deletion/duplication], EPCAM and GREM1 [deletion/duplication only]	Investigational
CPT®	0130U	Hereditary colon cancer disorders (eg, Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis), targeted mRNA sequence analysis panel (APC, CDH1, CHEK2, MLH1, MSH2, MSH6, MUTYH, PMS2, PTEN, and TP53) (List separately in addition to code for primary procedure)	Investigational
CPT®	0400U	Obstetrics (expanded carrier screening), 145 genes by next-generation sequencing, fragment analysis and multiplex ligation-dependent probe amplification, DNA, reported as carrier positive or negative	Investigational

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